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UNITED STATES PATENT APPLICATION

OF

Paul O. Sheppard, Stephen R. Jaspers, Zeren Gao

FOR

CONNECTIVE TISSUE GROWTH FACTOR HOMOLOGS

PATENT APPLICATION

97-75

Description

5 CONNECTIVE TISSUE GROWTH FACTOR HOMOLOGS

REFERENCE TO RELATED APPLICATIONS

This application is related to U.S. Patent
10 Provisional Application 60/075,300, filed on February 20,
1998, for which claims of benefit are made under 35 U.S.C.
§ 119(e)(1).

BACKGROUND OF THE INVENTION

15 Connective tissue growth factor (CTGF) is a
growth factor expressed by endothelial and fibroblastic
cells. Two members of the CTGF family are known, CTGF
(U.S. Patent 5,585,270 and U.S. Patent 5,408,040) and
CTGF-2 (WO 96/01896), incorporated herein by reference.
20 CTGF belongs to a family of growth factors that includes
CTGF, CTGF-2, insulin binding proteins (IBP) 1 and 2.
These growth factors have a cysteine-rich motif and
several other structural protein domains in common, and
have been demonstrated to have a role in cell
25 proliferation, differentiation, and chemotaxis.

Studies have suggested that CTGF is
immunologically similar to PDGF, with PDGF antibodies from
both A and B chains binding to CTGF (Bradham et al., J. of
Cell Biol. 114 (6):1285-1294, 1991), and it has been
30 reported that biological activity of CTGF can be blocked
using these antibodies.

It has recently been demonstrated that porcine
CTGF isolated from uterus is mitogenic for fibroblasts and
smooth muscle cells but not endothelial cells. In
35 addition, a N-terminally truncated form of the protein was
active as well (Brigstock et al., J. Biol. Chem.
272:20275-20282, 1997). It has been suggested that this

protein could play a role in the growth and remodeling of the endometrium and placenta.

The CTGF family is believed to play a role in production of the extracellular matrix components, such as 5 collagen and fibronectin. Collagen and fibronectin are components of many connective tissues, e.g., ligaments, cartilage, tendons and vessel walls. Current therapy for ligament repair is limited to immobilising the damaged tissue, i.e., stapling, or replacing the damaged tissue 10 with synthetic or natural grafts. These tissues are notoriously difficult to heal, even in healthy individuals, and compositions that would improve recovery time would be very valuable.

The present invention is directed to novel 15 polypeptides and polynucleotides encoding the polypeptide that show predominantly high expression in testis, trachea and bone marrow, thus providing a new molecule for regulating growth, differentiation, chemotaxis and induction of specialized cell functions in these tissues.

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SUMMARY OF THE INVENTION

It is an object of the present invention to provide an isolated polynucleotide molecule comprising a polynucleotide sequence that encodes a connective tissue 25 growth factor homolog polypeptide that is at least 70% identical to the amino acid sequence as shown in SEQ ID NO: 2 from residue 24 to residue 354.

In another embodiment, the present invention provides for a polynucleotide molecule wherein the 30 polynucleotide molecule comprises a region having the following motif as shown in SEQ ID NO: 23:
Cx{8,10}CxCCxxCx{7}Cx{5,6}Cx{5,7}Cx{12,13}Cx{7,8}Cx{20}
CxCx{6} Cx{12,14}Cx{13,17}C

wherein x{ } is the number of amino acid 35 residues between cysteines (C).

In another embodiment, the polynucleotide is 80% or 90% identical to the amino acid sequence as shown in SEQ ID NO: 2 from residue 24 to residue 354.

It is also the object of the present invention 5 to provide an isolated polynucleotide acid molecule that encodes a connective tissue growth factor homolog polypeptide, wherein the polynucleotide molecule is selected from the group consisting of (a) a molecules having the nucleotide sequence of SEQ ID NO:1 from 10 nucleotide 17 or 86 to nucleotide 1078, (b) a molecule encoding the amino acid sequence of SEQ ID NO:3 from nucleotide 1 or 70 to nucleotide 1062, and (c) a molecule that hybridizes under stringent wash conditions to a 15 polynucleotide molecule having the nucleotide sequence of nucleotides 86 to 1078 of SEQ ID NO:1, or the complement of nucleotides 86 to 1078 of SEQ ID NO:1.

In another embodiment, the differences in the amino acid sequence encoded by the polynucleotide and SEQ ID NO: 2 are conservative amino acid changes.

20 In other aspects, the present invention provides an expression vector comprising the following operably linked elements: a transcription promoter; a DNA segment comprising the isolated polynucleotide sequence that encodes a connective tissue growth factor homolog 25 polypeptide that is at least 70% identical to the amino acid sequence as shown in SEQ ID NO: 2 from residue 24 to residue 354; and a transcription terminator and a cultured host cell into which has been introduced the expression vector.

30 In another embodiment, the present invention provides a method of producing a connective tissue growth factor homolog polypeptide comprising: (a) culturing the host cells of expressing the CTGF homolog polypeptide; and (b) isolating the connective tissue growth factor homolog 35 polypeptide from the cultured host cells.

In another aspect, the present invention provides an isolated connective tissue growth factor

polypeptide comprising an amino acid sequence that is at least 70% identical to the amino acid sequence as shown in SEQ ID NO: 2 from residue 24 to residue 354.

In other embodiments, the present invention 5 provides CTGF homolog polypeptides wherein the amino acid sequence is at least 80% or 90% identical.

In another embodiment, the CTGF homolog polypeptide molecule comprises a region having the following motif as shown in SEQ ID NO: 23:
10 Cx{8,10}CxCCxxCx{7}Cx{5,6}Cx{5,7}Cx{12,13}Cx{7,8}Cx{20}
CxCx{6}Cx{12,14}Cx{13,17}C

wherein x{ } is the number of amino acid residues between cysteines (C).

In another aspect, the present invention 15 provides an antibody or antibody fragment that specifically binds with the CTGF homolog polypeptide.

In another aspect, the present invention provides a method of detecting the presence of connective tissue growth factor homolog polypeptide in a biological 20 sample, comprising the steps of: (a) contacting the biological sample with an antibody, or an antibody fragment of claim 14, wherein the contacting is performed under conditions that allow the binding of the antibody or antibody fragment to the biological sample, and (b) 25 detecting any of the bound antibody or bound antibody fragment.

In another aspect, the present invention provides an anti-idiotype antibody, or anti-idiotype antibody fragment, that specifically binds with the 30 antibody or antibody fragment.

In another aspect, the present invention provides a method of detecting a chromosome 6q abnormality in sample from an individual comprising: (a) obtaining zCTGF4 RNA from the sample; (b) generating zCTGF4 cDNA by 35 polymerase chain reaction; and (c) comparing the nucleic acid sequence of the zCTGF4 cDNA to the nucleic acid sequence as shown in SEQ ID NO: 1.

In another embodiment, the present invention provides that the difference between the sequence of the zCTGF4 cDNA or zCTGF4 gene in the sample and the zCTGF4 sequence as shown in SEQ ID NO: 1 is indicative of 5 chromosome 6q abnormality.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 illustrates a multiple alignment of human zCTGF4 (SEQ ID NO: 2) and other members of the 10 connective tissue growth factor family that includes, mouse zCTGF2 (SEQ ID NO: 5), human NOV (NOV HU; SEQ ID NO: 25), human CTGF1 (CTGF H; SEQ ID NO: 26), human 15 insulin binding protein 1 (IBP1 H; SEQ ID NO: 27), and human insulin binding protein 2 (IBP2 H; SEQ ID NO: 28).

DETAILED DESCRIPTION OF THE INVENTION

Prior to setting forth the invention in detail, it may be helpful to the understanding thereof to define the following terms:

The term "affinity tag" is used herein to denote 20 a polypeptide segment that can be attached to a second polypeptide to provide for purification or detection of the second polypeptide or provide sites for attachment of the second polypeptide to a substrate. In principal, any 25 peptide or protein for which an antibody or other specific binding agent is available can be used as an affinity tag. Affinity tags include a poly-histidine tract, protein A (Nilsson et al., EMBO J. 4:1075, 1985; Nilsson et al., Methods Enzymol. 198:3, 1991), glutathione S transferase 30 (Smith and Johnson, Gene 67:31, 1988), Glu-Glu affinity tag (Grussenmeyer et al., Proc. Natl. Acad. Sci. USA 82:7952-4, 1985), substance P, FlagTM peptide (Hopp et al., Biotechnology 6:1204-10, 1988), streptavidin binding peptide, or other antigenic epitope or binding domain. 35 See, in general, Ford et al., Protein Expression and Purification 2: 95-107, 1991. DNAs encoding affinity tags

are available from commercial suppliers (e.g., Pharmacia Biotech, Piscataway, NJ).

The term "allelic variant" is used herein to denote any of two or more alternative forms of a gene occupying the same chromosomal locus. Allelic variation arises naturally through mutation, and may result in phenotypic polymorphism within populations. Gene mutations can be silent (no change in the encoded polypeptide) or may encode polypeptides having altered amino acid sequence. The term allelic variant is also used herein to denote a protein encoded by an allelic variant of a gene.

The terms "amino-terminal" and "carboxyl-terminal" are used herein to denote positions within polypeptides. Where the context allows, these terms are used with reference to a particular sequence or portion of a polypeptide to denote proximity or relative position. For example, a certain sequence positioned carboxyl-terminal to a reference sequence within a polypeptide is located proximal to the carboxyl terminus of the reference sequence, but is not necessarily at the carboxyl terminus of the complete polypeptide.

The term "complement/anti-complement pair" denotes non-identical moieties that form a non-covalently associated, stable pair under appropriate conditions. For instance, biotin and avidin (or streptavidin) are prototypical members of a complement/anti-complement pair. Other exemplary complement/anti-complement pairs include receptor/ligand pairs, antibody/antigen (or hapten or epitope) pairs, sense/antisense polynucleotide pairs, and the like. Where subsequent dissociation of the complement/anti-complement pair is desirable, the complement/anti-complement pair preferably has a binding affinity of $<10^9 \text{ M}^{-1}$.

The term "complements of a polynucleotide molecule" denotes a polynucleotide molecule having a complementary base sequence and reverse orientation as

compared to a reference sequence. For example, the sequence 5' ATGCACGGG 3' is complementary to 5' CCCGTGCAT 3'.

The term "contig" denotes a polynucleotide that 5 has a contiguous stretch of identical or complementary sequence to another polynucleotide. Contiguous sequences are said to "overlap" a given stretch of polynucleotide sequence either in their entirety or along a partial stretch of the polynucleotide.

10 The term "degenerate nucleotide sequence" denotes a sequence of nucleotides that includes one or more degenerate codons (as compared to a reference polynucleotide molecule that encodes a polypeptide). Degenerate codons contain different triplets of 15 nucleotides, but encode the same amino acid residue (i.e., GAU and GAC triplets each encode Asp).

The term "expression vector" is used to denote a DNA molecule, linear or circular, that comprises a segment encoding a polypeptide of interest operably linked to 20 additional segments that provide for its transcription. Such additional segments include promoter and terminator sequences, and may also include one or more origins of replication, one or more selectable markers, an enhancer, a polyadenylation signal, etc. Expression vectors are 25 generally derived from plasmid or viral DNA, or may contain elements of both.

The term "isolated", when applied to a polynucleotide, denotes that the polynucleotide has been removed from its natural genetic milieu and is thus free 30 of other extraneous or unwanted coding sequences, and is in a form suitable for use within genetically engineered protein production systems. Such isolated molecules are those that are separated from their natural environment and include cDNA and genomic clones. Isolated DNA 35 molecules of the present invention are free of other genes with which they are ordinarily associated, but may include naturally occurring 5' and 3' untranslated regions such as

promoters and terminators. The identification of associated regions will be evident to one of ordinary skill in the art (see for example, Dynan and Tijan, Nature 316:774-78, 1985).

5 An "isolated" polypeptide or protein is a polypeptide or protein that is found in a condition other than its native environment, such as apart from blood and animal tissue. In a preferred form, the isolated polypeptide is substantially free of other polypeptides, 10 particularly other polypeptides of animal origin. It is preferred to provide the polypeptides in a highly purified form, i.e. greater than 95% pure, more preferably greater than 99% pure. When used in this context, the term "isolated" does not exclude the presence of the same 15 polypeptide in alternative physical forms, such as dimers or alternatively glycosylated or derivatized forms.

The term "operably linked", when referring to DNA segments, indicates that the segments are arranged so that they function in concert for their intended purposes, 20 e.g., transcription initiates in the promoter and proceeds through the coding segment to the terminator.

The term "ortholog" denotes a polypeptide or protein obtained from one species that is the functional counterpart of a polypeptide or protein from a different 25 species. Sequence differences among orthologs are the result of speciation.

"Paralogs" are distinct but structurally related proteins made by an organism. Paralogs are believed to arise through gene duplication. For example, α -globin, β -globin, and myoglobin are paralogs of each other.

A "polynucleotide" is a single- or double-stranded polymer of deoxyribonucleotide or ribonucleotide bases read from the 5' to the 3' end. Polynucleotides include RNA and DNA, and may be isolated from natural 35 sources, synthesized *in vitro*, or prepared from a combination of natural and synthetic molecules. Sizes of

polynucleotides are expressed as base pairs (abbreviated "bp"), nucleotides ("nt"), or kilobases ("kb"). Where the context allows, the latter two terms may describe polynucleotides that are single-stranded or double-stranded. When the term is applied to double-stranded molecules it is used to denote overall length and will be understood to be equivalent to the term "base pairs". It will be recognized by those skilled in the art that the two strands of a double-stranded polynucleotide may differ slightly in length and that the ends thereof may be staggered as a result of enzymatic cleavage; thus all nucleotides within a double-stranded polynucleotide molecule may not be paired. Such unpaired ends will in general not exceed 20 nt in length.

A "polypeptide" is a polymer of amino acid residues joined by peptide bonds, whether produced naturally or synthetically. Polypeptides of less than about 10 amino acid residues are commonly referred to as "peptides".

The term "promoter" is used herein for its art-recognized meaning to denote a portion of a gene containing DNA sequences that provide for the binding of RNA polymerase and initiation of transcription. Promoter sequences are commonly, but not always, found in the 5' non-coding regions of genes.

A "protein" is a macromolecule comprising one or more polypeptide chains. A protein may also comprise non-peptidic components, such as carbohydrate groups. Carbohydrates and other non-peptidic substituents may be added to a protein by the cell in which the protein is produced, and will vary with the type of cell. Proteins are defined herein in terms of their amino acid backbone structures; substituents such as carbohydrate groups are generally not specified, but may be present nonetheless.

The term "receptor" denotes a cell-associated protein that binds to a bioactive molecule (i.e., a ligand) and mediates the effect of the ligand on the cell.

Membrane-bound receptors are characterized by a multi-peptide structure comprising an extracellular ligand-binding domain and an intracellular effector domain that is typically involved in signal transduction. Binding of 5 ligand to receptor results in a conformational change in the receptor that causes an interaction between the effector domain and other molecule(s) in the cell. This interaction in turn leads to an alteration in the metabolism of the cell. Metabolic events that are linked 10 to receptor-ligand interactions include gene transcription, phosphorylation, dephosphorylation, increases in cyclic AMP production, mobilization of cellular calcium, mobilization of membrane lipids, cell adhesion, hydrolysis of inositol lipids and hydrolysis of 15 phospholipids. In general, receptors can be membrane bound, cytosolic or nuclear; monomeric (e.g., thyroid stimulating hormone receptor, beta-adrenergic receptor) or multimeric (e.g., PDGF receptor, growth hormone receptor, IL-3 receptor, GM-CSF receptor, G-CSF receptor, 20 erythropoietin receptor and IL-6 receptor).

The term "secretory signal sequence" denotes a DNA sequence that encodes a polypeptide (a "secretory peptide") that, as a component of a larger polypeptide, directs the larger polypeptide through a secretory pathway 25 of a cell in which it is synthesized. The larger polypeptide is commonly cleaved to remove the secretory peptide during transit through the secretory pathway.

The term "splice variant" is used herein to denote alternative forms of RNA transcribed from a gene. 30 Splice variation arises naturally through use of alternative splicing sites within a transcribed RNA molecule, or less commonly between separately transcribed RNA molecules, and may result in several mRNAs transcribed from the same gene. Splice variants may encode 35 polypeptides having altered amino acid sequence. The term splice variant is also used herein to denote a protein

encoded by a splice variant of an mRNA transcribed from a gene.

Molecular weights and lengths of polymers determined by imprecise analytical methods (e.g., gel 5 electrophoresis) will be understood to be approximate values. When such a value is expressed as "about" X or "approximately" X, the stated value of X will be understood to be accurate to $\pm 10\%$.

All references cited herein are incorporated by 10 reference in their entirety.

The present invention is based in part upon the discovery of a novel DNA sequence that encodes a connective tissue growth factor homolog polypeptide, i.e., 15 polypeptides having homology to other members of a family of growth factors that are secreted and contain a cysteine motif of the formula:

$Cx\{8,10\}CxCCxxCx\{7\}Cx\{5,6\}Cx\{5,7\}Cx\{12,13\}Cx\{7,8\}Cx\{20\}$
 $CxCx\{6\}Cx\{12,14\}Cx\{13,17\}C$

20 wherein x{ } is the number of amino acid residues between cysteines (C) as shown in SEQ ID NO: 23. This motif represents a consecutive domain arrangement known as a insulin-like growth factor binding domain followed by a Von Willebrand Factor c (VWFc) domain (Bork, 25 FEBS Letts. 327:125-130, 1993). The protein domain arrangement is unique to the CTGF/NOV protein structure family. The insulin-like growth factor binding domain is represented by the motif:

$Cx\{8,10\}CxCCxxCx\{7\}Cx\{5,6\}Cx\{5,7\}$

30 wherein x{ } is the number of amino acid residues between cysteines (C) as shown in SEQ ID NO: 24. This pattern is found in all known members of the connective tissue growth factors and insulin binding protein family (for example: human CTGF, human CTGF-2, 35 human NOV, human IBPs 1 and 2), and is unique to these proteins.

Analysis of the tissue distribution of the mRNA corresponding to this novel DNA showed that expression was highest in testis, followed by apparent but decreased expression levels in trachea, bone marrow and kidney 5 tissue. The connective tissue growth factor homolog polypeptide has been designated zCTGF4.

The ortholog from mouse has also been identified and designated zCTGF2. A DNA sequence and corresponding putative amino acid sequence are shown in SEQ ID NOS: 4 10 and 5, respectively.

The novel zCTGF4 polypeptides of the present invention were initially identified by querying an EST database for homologous sequences to connective tissue growth factor and insulin-binding proteins. A single EST 15 sequence was discovered and predicted to be related to the connective tissue growth factor family. Sequence analysis of the clone from which the EST was derived revealed that the clone contained a defective cDNA with incorrect sequence. Isolation of an independent clone from a testis 20 library revealed that the original cDNA clone contained intronic sequence at the 5' end of the EST. The nucleotide sequence is described in SEQ ID NO. 1 from nucleotide 17 to nucleotide 1078, and its deduced amino acid sequence is described in SEQ ID NO. 2.

Analysis of the DNA encoding a zCTGF4 polypeptide (SEQ ID NO: 1) revealed an open reading frame encoding 354 amino acids (SEQ ID NO: 2) comprising a secretory signal peptide of 23 amino acid residues (residue 1 (Met) to residue 23 (Gly) of SEQ ID NO: 2) and 30 a mature polypeptide of 331 amino acids (residue 24 (Thr) to residue 354 (Leu) of SEQ ID NO: 2). An alternatively spliced variant has a 16 amino acid secretory signal peptide (residue 1 (Met) to residue 16 (Gly) of SEQ ID NO: 2, and a mature polypeptide of 338 amino acid residues 35 (residue 17 (Phe) to residue 354 (Leu) of SEQ ID NO: 2.

CTGF family members are characterized by a multi-domain structure comprising a IBP domain (amino acid

residue 59 (Pro) to 102 (Tyr) of SEQ ID NO: 2) that has been suggested as an insulin growth factor binding domain (Kiefer et al., J. Biol. Chem. 266:9043-9049, 1991), a von Willebrand factor c domain (amino acid residue 114 (Cys) 5 to 179 (Cys) of SEQ ID NO: 2) that may be involved in multimerization, a variable domain (amino acid 180 (Ser) to 208 (Lys), that may be involved in tissue, matrix, growth factor or receptor specific interactions; and a sulfated glycoconjugate binding motif domain (amino acid 10 residue 209 (Cys) to 252 (Cys) of SEQ ID NO: 2) that is thought to be involved in binding large macromolecules (Holt et al., J. Biol. Chem. 265:2852-2855, 1990).

CTGF has recently been demonstrated to bind IGF at low affinity (Kim et al., Proc. Nat. Acad. Sci. 94:12981-12986, 1997), and thereby expands the insulin-like growth factor binding protein (IGFBP) superfamily to include the CTGFS. The superfamily of IGFBPs now comprises both proteins that bind IGF high affinity (e.g., IGFBP 1-5) and proteins that bind IGF with low affinity (nov, cyr61 and CTGF), suggesting proteins affect cell growth in both IGF-dependent and IGF-independent manners. The IGF binding potential of members of the CTGF family, including zCTGF4, may act as a competitive inhibitor of the biologically free component of IGF. ZCTGF4 may affect other pathways that IGFBPs play a role in, for example, competing with endogenous IGFBP for proteases that degrade the IGF/IGFBP complex, resulting in changes in the circulating levels of IGF.

A multiple alignment, as shown in Figure 1, also 30 revealed that zCTGF4, like several other known members of the CTGF family, has a heparin binding domain that has been suggested as a receptor binding and dimerization domain. The heparin binding domain is shown in SEQ ID NO: 2 from residue 262 (Ile) to residue 295 (Phe). The domain 35 structure has been reviewed by Brigstock et al., *ibid.*, 1997 and Bork, FEBS Letts. 327:125-130, 1993, both incorporated herein by reference.

It is generally believed that under selective pressure for organisms to acquire new biological functions, new CTGF family members arose from duplication of existing genes leading to the existence of multi-gene families. Family members thus contain vestiges of the ancestral gene, and these characteristic features can be exploited in the isolation and identification of additional family members. It has recently been demonstrated that N-terminally truncated CTGF molecules isolated from uterine secretory fluids have mitogenic activity and will bind heparin (Brigstock et al., *ibid.*, 1997), and may have activity. An N-terminally truncated zCTGF4 molecule would be expected to have similar activity, and would comprise molecules without the IBP domain and may or may not be C-terminally truncated as well.

SEQ ID NO: 3 is a degenerate polynucleotide sequence that encompasses all polynucleotides that could encode the zCTGF4 polypeptide of SEQ ID NO: 2 (amino acids 1 or 24 to 354). Thus, zCTGF4 polypeptide-encoding polynucleotides ranging from nucleotide 17 or 85 to nucleotide 1078 of SEQ ID NO: 2 or nucleotide 1 or 69 to 1062 of SEQ ID NO: 3 are contemplated by the present invention. Also contemplated by the present invention are fragments and fusions as described herein with respect to SEQ ID NO: 1, which are formed from analogous regions of SEQ ID NO: 3, wherein nucleotides 191 to 322 of SEQ ID NO: 1 correspond to nucleotides 175 to 306 of SEQ ID NO: 3, for the IBP domain; wherein nucleotides 356 to 553 of SEQ ID NO: 1 correspond to nucleotides 340 to 537 of SEQ ID NO: 3, for the von Willebrand factor (VWFc) domain; wherein nucleotides 554 to 640 of SEQ ID NO: 1 correspond to nucleotides 538 to 624 of SEQ ID NO: 3 for the variable region; wherein nucleotides 641 to 772 of SEQ ID NO: 1 correspond to nucleotides 625 to 756 of SEQ ID NO: 3, for the sulfated glycoconjugate binding domain; and wherein nucleotide 800 to nucleotide 904 of SEQ ID NO: 1

correspond to nucleotide 784 to nucleotide 888 of SEQ ID NO: 3 for the heparin binding domain. Table 1 sets forth the one-letter codes used within SEQ ID NO: 3 to denote degenerate nucleotide positions. "Resolutions" are the
5 nucleotides denoted by a code letter. "Complement" indicates the code for the complementary nucleotide(s). For example, the code Y denotes either C or T, and its complement R denotes A or G, A being complementary to T, and G being complementary to C.

TABLE 1

Nucleotide	Resolution	Complement	Resolution
A	A	T	T
C	C	G	G
G	G	C	C
T	T	A	A
R	A G	Y	C T
Y	C T	R	A G
M	A C	K	G T
K	G T	M	A C
S	C G	S	C G
W	A T	W	A T
H	A C T	D	A G T
B	C G T	V	A C G
V	A C G	B	C G T
D	A G T	H	A C T
N	A C G T	N	A C G T

The degenerate codons used in SEQ ID NOS: 2 and
 5 5, encompassing all possible codons for a given amino acid, are set forth in Table 2.

TABLE 2

Amino Acid	Letter Code	One Codons	Degenerate Codon
Cys	C	TGC TGT	TGY
Ser	S	AGC AGT TCA TCC TCG TCT	WSN
Thr	T	ACA ACC ACG ACT	ACN
Pro	P	CCA CCC CCG CCT	CCN
Ala	A	GCA GCC GCG GCT	GCN
Gly	G	GGA GGC GGG GGT	GGN
Asn	N	AAC AAT	AAY
Asp	D	GAC GAT	GAY
Glu	E	GAA GAG	GAR
Gln	Q	CAA CAG	CAR
His	H	CAC CAT	CAY
Arg	R	AGA AGG CGA CGC CGG CGT	MGN
Lys	K	AAA AAG	AAR
Met	M	ATG	ATG
Ile	I	ATA ATC ATT	ATH
Leu	L	CTA CTC CTG CTT TTA TTG	YTN
Val	V	GTA GTC GTG GTT	GTN
Phe	F	TTC TTT	TTY
Tyr	Y	TAC TAT	TAY
Trp	W	TGG	TGG
Ter	.	TAA TAG TGA	TRR
Asn Asp	B		RAY
Glu Gln	Z		SAR
Any	X		NNN

One of ordinary skill in the art will appreciate that some ambiguity is introduced in determining a degenerate codon, representative of all possible codons encoding each amino acid. For example, the degenerate 5 codon for serine (WSN) can, in some circumstances, encode arginine (AGR), and the degenerate codon for arginine (MGN) can, in some circumstances, encode serine (AGY). A similar relationship exists between codons encoding phenylalanine and leucine. Thus, some polynucleotides 10 encompassed by the degenerate sequence may encode variant amino acid sequences, but one of ordinary skill in the art can easily identify such variant sequences by reference to the amino acid sequence of SEQ ID NOS: 2 and 5. Variant sequences can be readily tested for functionality as 15 described herein.

One of ordinary skill in the art will also appreciate that different species can exhibit "preferential codon usage." In general, see, Grantham, et al., Nuc. Acids Res., 8:1893-912, 1980; Haas, et al. Curr. Biol., 6:315-24, 1996; Wain-Hobson, et al., Gene, 13:355-20 64, 1981; Grosjean and Fiers, Gene, 18:199-209, 1982; Holm, Nuc. Acids Res., 14:3075-87, 1986; Ikemura, J. Mol. Biol., 158:573-97, 1982. As used herein, the term 25 "preferential codon usage" or "preferential codons" is a term of art referring to protein translation codons that are most frequently used in cells of a certain species, thus favoring one or a few representatives of the possible codons encoding each amino acid (See Table 2). For example, the amino acid Threonine (Thr) may be encoded by 30 ACA, ACC, ACG, or ACT, but in mammalian cells ACC is the most commonly used codon; in other species, for example, insect cells, yeast, viruses or bacteria, different Thr codons may be preferential. Preferential codons for a particular species can be introduced into the 35 polynucleotides of the present invention by a variety of methods known in the art. Introduction of preferential codon sequences into recombinant DNA can, for example,

enhance production of the protein by making protein translation more efficient within a particular cell type or species. Therefore, the degenerate codon sequence disclosed in SEQ ID NO: 3 serves as a template for 5 optimizing expression of polynucleotides in various cell types and species commonly used in the art and disclosed herein. Sequences containing preferential codons can be tested and optimized for expression in various species, and tested for functionality as disclosed herein.

10 The present invention further provides variant polypeptides and nucleic acid molecules that represent counterparts from other species (orthologs). These species include, but are not limited to mammalian, avian, amphibian, reptile, fish, insect and other vertebrate and 15 invertebrate species. Of particular interest are zCTGF4 polypeptides from other mammalian species, including murine, porcine, ovine, bovine, canine, feline, equine, and other primate polypeptides. Orthologs of human zCTGF4 can be cloned using information and compositions provided 20 by the present invention in combination with conventional cloning techniques. For example, a cDNA can be cloned using mRNA obtained from a tissue or cell type that expresses zCTGF4 as disclosed herein. Suitable sources of 25 mRNA can be identified by probing northern blots with probes designed from the sequences disclosed herein. A library is then prepared from mRNA of a positive tissue or cell line. The mouse sequence ZCTGF2 is a representative ortholog of the human connective tissue growth factor, zCTGF4, and is disclosed herein as SEQ ID NOS: 4 and 5.

30 An zCTGF4-encoding cDNA can then be isolated by a variety of methods, such as by probing with a complete or partial human cDNA or with one or more sets of degenerate probes based on the disclosed sequences. A cDNA can also be cloned using the polymerase chain 35 reaction with primers designed from the representative human zCTGF4 sequences disclosed herein. Within an additional method, the cDNA library can be used to

transform or transfect host cells, and expression of the cDNA of interest can be detected with an antibody to zCTGF4 polypeptide. Similar techniques can also be applied to the isolation of genomic clones.

5 The present invention provides polynucleotide molecules including DNA and RNA molecules that encode the zCTGF4 polypeptides disclosed above.

ZCTGF4 polynucleotide sequences disclosed herein can also be used as probes or primers to clone 5' non-
10 coding regions of a zCTGF4 gene. In view of the tissue-specific expression observed for zCTGF4 by Northern blotting, this gene region is expected to provide for testis-, trachea-, bone marrow-, and kidney-specific expression. Promoter elements from a zCTGF4 gene could
15 thus be used to direct the tissue-specific expression of heterologous genes in, for example, transgenic animals or patients treated with gene therapy. Cloning of 5' flanking sequences also facilitates production of zCTGF4 proteins by "gene activation" as disclosed in U.S. Patent
20 No. 5,641,670. Briefly, expression of an endogenous zCTGF4 gene in a cell is altered by introducing into the zCTGF4 locus a DNA construct comprising at least a targeting sequence, a regulatory sequence, an exon, and an unpaired splice donor site. The targeting sequence is a
25 zCTGF4 5' non-coding sequence that permits homologous recombination of the construct with the endogenous zCTGF4 locus, whereby the sequences within the construct become operably linked with the endogenous zCTGF4 coding sequence. In this way, an endogenous zCTGF4 promoter can
30 be replaced or supplemented with other regulatory sequences to provide enhanced, tissue-specific, or otherwise regulated expression.

Those skilled in the art will recognize that the sequence disclosed in SEQ ID NO:1 represents a single allele of human zCTGF4 and that allelic variation and alternative splicing are expected to occur. Allelic variants of this sequence can be cloned by probing cDNA or

genomic libraries from different individuals according to standard procedures. Allelic variants of the nucleotide sequence shown in SEQ ID NO:1, including those containing silent mutations and those in which mutations result in 5 amino acid sequence changes, are within the scope of the present invention, as are proteins which are allelic variants of SEQ ID NO:2. cDNA molecules generated from alternatively spliced mRNAs, which retain the properties of the zCTGF4 polypeptide are included within the scope of 10 the present invention, as are polypeptides encoded by such cDNAs and mRNAs. Allelic variants and splice variants of these sequences can be cloned by probing cDNA or genomic libraries from different individuals or tissues according to standard procedures known in the art.

15 The present invention provides methods for using zCTGF4 polynucleotides and polypeptides to diagnose chromosomal disorders associated with abnormal expression of the zCTGF4 protein. Detectable chromosomal mutations at the zCTGF4 gene locus include, but are not limited to, 20 aneuploidy, gene copy number changes, insertions, deletions, restriction site changes and rearrangements. Such aberrations can be identified by employing molecular genetic techniques, such as restriction fragment length polymorphism (RFLP) analysis, short tandem repeat (STR) 25 analysis employing PCR techniques, and other genetic linkage analysis techniques known in the art (*Molecular Cloning: A Laboratory Manual*, 2nd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989, and Ausubel et al., eds., *Current Protocols in Molecular 30 Biology*, John Wiley and Sons, Inc., NY, 1987; A.J. Marian, *Chest* 108:255-65, 1995). Analyses of DNA samples can detect deletions and insertions by changes in size in amplified DNA products by comparing a sample DNA to a normal zCTGF4 DNA standard. Mismatches in duplex DNA can 35 be detected by RNase digestion or differences in melting temperature. Other methods for detecting differences in sequences include, changes electrophoretic motility,

Southern analysis, and direct DNA sequencing. Recently, techniques for accessing genetic information with high-density arrays have been available (Chee et al., Science 274:610-614, 1996), and can analyze large fragments of genomic DNA with high resolution.

Analysis of chromosomal DNA using the zCTGF4 polynucleotide sequence is useful for correlating disease with abnormalities localized to chromosome 6. The zCTGF4 gene has been localized to chromosome 6q22.1. Studies of the DNA sequences, cDNA and/or genomic DNA, of some of the individuals presenting disease that contain a mutation in the sequence of the zCTGF4 gene, that is not present in normal individuals, can provide strong evidence for the mutation as causative of the disease. In one embodiment, the methods of the present invention provide a method of detecting a chromosome 6q abnormality in sample from an individual comprising: (a) obtaining zCTGF4 RNA from the sample; (b) generating zCTGF4 cDNA by polymerase chain reaction; and (c) comparing the nucleic acid sequence of the zCTGF4 cDNA to the nucleic acid sequence as shown in SEQ ID NO: 1. In further embodiments, the difference between the sequence of the zCTGF4 cDNA or zCTGF4 gene in the sample and the zCTGF4 sequence as shown in SEQ ID NO: 1 is indicative of chromosome 6q abnormality.

Within preferred embodiments of the invention, the isolated nucleic acid molecules can hybridize under stringent conditions to nucleic acid molecules having the nucleotide sequence of SEQ ID NO:1, to nucleic acid molecules having the nucleotide sequence of nucleotides 86 to 1078 of SEQ ID NO:1, or to nucleic acid molecules having a nucleotide sequence complementary to SEQ ID NO:1. In general, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe.

A pair of nucleic acid molecules, such as DNA-DNA, RNA-RNA and DNA-RNA, can hybridize if the nucleotide sequences have some degree of complementarity. Hybrids can tolerate mismatched base pairs in the double helix, but the stability of the hybrid is influenced by the degree of mismatch. The T_m of the mismatched hybrid decreases by 1°C for every 1-1.5% base pair mismatch. Varying the stringency of the hybridization conditions allows control over the degree of mismatch that will be present in the hybrid. The degree of stringency increases as the hybridization temperature increases and the ionic strength of the hybridization buffer decreases. Stringent hybridization conditions encompass temperatures of about 5-25°C below the T_m of the hybrid and a hybridization buffer having up to 1 M Na⁺. Higher degrees of stringency at lower temperatures can be achieved with the addition of formamide which reduces the T_m of the hybrid about 1°C for each 1% formamide in the buffer solution. Generally, such stringent conditions include temperatures of 20-70°C and a hybridization buffer containing up to 6x SSC and 0-50% formamide. A higher degree of stringency can be achieved at temperatures of from 40-70°C with a hybridization buffer having up to 4x SSC and from 0-50% formamide. Highly stringent conditions typically encompass temperatures of 42-70°C with a hybridization buffer having up to 1x SSC and 0-50% formamide. Different degrees of stringency can be used during hybridization and washing to achieve maximum specific binding to the target sequence. Typically, the washes following hybridization are performed at increasing degrees of stringency to remove non-hybridized polynucleotide probes from hybridized complexes.

The above conditions are meant to serve as a guide and it is well within the abilities of one skilled in the art to adapt these conditions for use with a particular polypeptide hybrid. The T_m for a specific target sequence is the temperature (under defined

conditions) at which 50% of the target sequence will hybridize to a perfectly matched probe sequence. Those conditions which influence the T_m include, the size and base pair content of the polynucleotide probe, the ionic strength of the hybridization solution, and the presence of destabilizing agents in the hybridization solution. Numerous equations for calculating T_m are known in the art, and are specific for DNA, RNA and DNA-RNA hybrids and polynucleotide probe sequences of varying length (see, for example, Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, Second Edition (Cold Spring Harbor Press 1989); Ausubel *et al.*, (eds.), *Current Protocols in Molecular Biology* (John Wiley and Sons, Inc. 1987); Berger and Kimmel (eds.), *Guide to Molecular Cloning Techniques*, (Academic Press, Inc. 1987); and Wetmur, *Crit. Rev. Biochem. Mol. Biol.* 26:227 (1990)). Sequence analysis software such as OLIGO 6.0 (LSR; Long Lake, MN) and Primer Premier 4.0 (Premier Biosoft International; Palo Alto, CA), as well as sites on the Internet, are available tools for analyzing a given sequence and calculating T_m based on user defined criteria. Such programs can also analyze a given sequence under defined conditions and identify suitable probe sequences. Typically, hybridization of longer polynucleotide sequences, >50 base pairs, is performed at temperatures of about 20-25°C below the calculated T_m . For smaller probes, <50 base pairs, hybridization is typically carried out at the T_m or 5-10°C below. This allows for the maximum rate of hybridization for DNA-DNA and DNA-RNA hybrids.

The length of the polynucleotide sequence influences the rate and stability of hybrid formation. Smaller probe sequences, <50 base pairs, reach equilibrium with complementary sequences rapidly, but may form less stable hybrids. Incubation times of anywhere from minutes to hours can be used to achieve hybrid formation. Longer probe sequences come to equilibrium more slowly, but form more stable complexes even at lower temperatures.

Incubations are allowed to proceed overnight or longer. Generally, incubations are carried out for a period equal to three times the calculated Cot time. Cot time, the time it takes for the polynucleotide sequences to 5 reassociate, can be calculated for a particular sequence by methods known in the art.

The base pair composition of polynucleotide sequence will effect the thermal stability of the hybrid complex, thereby influencing the choice of hybridization 10 temperature and the ionic strength of the hybridization buffer. A-T pairs are less stable than G-C pairs in aqueous solutions containing sodium chloride. Therefore, the higher the G-C content, the more stable the hybrid. Even distribution of G and C residues within the sequence 15 also contribute positively to hybrid stability. In addition, the base pair composition can be manipulated to alter the T_m of a given sequence. For example, 5-methyldeoxycytidine can be substituted for deoxycytidine and 5-bromodeoxuridine can be substituted for thymidine to 20 increase the T_m , whereas 7-deaz-2'-deoxyguanosine can be substituted for guanosine to reduce dependence on T_m .

The ionic concentration of the hybridization buffer also affects the stability of the hybrid. Hybridization buffers generally contain blocking agents 25 such as Denhardt's solution (Sigma Chemical Co., St. Louis, Mo.), denatured salmon sperm DNA, tRNA, milk powders (BLOTO), heparin or SDS, and a Na^+ source, such as SSC (1x SSC: 0.15 M sodium chloride, 15 mM sodium citrate) or SSPE (1x SSPE: 1.8 M NaCl, 10 mM NaH_2PO_4 , 1 mM EDTA, pH 30 7.7). By decreasing the ionic concentration of the buffer, the stability of the hybrid is increased. Typically, hybridization buffers contain from between 10 mM - 1 M Na^+ . The addition of destabilizing or denaturing 35 agents such as formamide, tetraalkylammonium salts, guanidinium cations or thiocyanate cations to the hybridization solution will alter the T_m of a hybrid. Typically, formamide is used at a concentration of up to

50% to allow incubations to be carried out at more convenient and lower temperatures. Formamide also acts to reduce non-specific background when using RNA probes.

As an illustration, a nucleic acid molecule encoding a variant zCTGF4 polypeptide can be hybridized with a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1 (or its complement) at 42°C overnight in a solution comprising 50% formamide, 5xSSC (1xSSC: 0.15 M sodium chloride and 15 mM sodium citrate), 50 mM sodium phosphate (pH 7.6), 5x Denhardt's solution (100x Denhardt's solution: 2% (w/v) Ficoll 400, 2% (w/v) polyvinylpyrrolidone, and 2% (w/v) bovine serum albumin), 10% dextran sulfate, and 20 µg/ml denatured, sheared salmon sperm DNA. One of skill in the art can devise variations of these hybridization conditions. For example, the hybridization mixture can be incubated at a higher temperature, such as about 65°C, in a solution that does not contain formamide. Moreover, premixed hybridization solutions are available (e.g., EXPRESSHYB Hybridization Solution from CLONTECH Laboratories, Inc.), and hybridization can be performed according to the manufacturer's instructions.

Following hybridization, the nucleic acid molecules can be washed to remove non-hybridized nucleic acid molecules under stringent conditions, or under highly stringent conditions. Typical stringent washing conditions include washing in a solution of 0.5x - 2x SSC with 0.1% sodium dodecyl sulfate (SDS) at 55 - 65°C. That is, nucleic acid molecules encoding a variant zCTGF4 polypeptide hybridize with a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1 (or its complement) under stringent washing conditions, in which the wash stringency is equivalent to 0.5x - 2x SSC with 0.1% SDS at 55 - 65°C, including 0.5x SSC with 0.1% SDS at 55°C, or 2xSSC with 0.1% SDS at 65°C. One of skill in the art can readily devise equivalent conditions, for example, by substituting SSPE for SSC in the wash solution.

Typical highly stringent washing conditions include washing in a solution of 0.1x - 0.2x SSC with 0.1% sodium dodecyl sulfate (SDS) at 50 - 65°C. In other words, nucleic acid molecules encoding a variant zCTGF4 polypeptide hybridize with a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1 (or its complement) under highly stringent washing conditions, in which the wash stringency is equivalent to 0.1x - 0.2x SSC with 0.1% SDS at 50 - 65°C, including 0.1x SSC with 0.1% SDS at 50°C, or 0.2xSSC with 0.1% SDS at 65°C.

The present invention also provides isolated zCTGF4 polypeptides that have a substantially similar sequence identity to the polypeptides of SEQ ID NO:2, or their orthologs. The term "substantially similar sequence identity" is used herein to denote polypeptides comprising at least 70%, at least 80%, at least 90%, at least 95% or greater than 95% sequence identity to the sequences shown in SEQ ID NO:2, or their orthologs. The present invention also includes polypeptides that comprise an amino acid sequence having at least 70%, at least 80%, at least 90%, at least 95% or greater than 95% sequence identity to the sequence of amino acid residues 1 or 24 to 354 of SEQ ID NO:2. The present invention further includes nucleic acid molecules that encode such polypeptides. Methods for determining percent identity are described below.

The present invention also contemplates zCTGF4 variant nucleic acid molecules that can be identified using two criteria: a determination of the similarity between the encoded polypeptide with the amino acid sequence of SEQ ID NO:2, and/or a hybridization assay, as described above. Such zCTGF4 variants include nucleic acid molecules (1) that hybridize with a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1 (or its complement) under stringent washing conditions, in which the wash stringency is equivalent to 0.5x - 2x SSC with 0.1% SDS at 55 - 65°C, or (2) that encode a polypeptide having at least 70%, at least 80%, at least

90%, at least 95% or greater than 95% sequence identity to the amino acid sequence of SEQ ID NO:2. Alternatively, zCTGF4 variants can be characterized as nucleic acid molecules (1) that hybridize with a nucleic acid molecule 5 having the nucleotide sequence of SEQ ID NO:1 (or its complement) under highly stringent washing conditions, in which the wash stringency is equivalent to 0.1x - 0.2x SSC with 0.1% SDS at 50 - 65°C, and (2) that encode a polypeptide having at least 70%, at least 80%, at least 10 90%, at least 95% or greater than 95% sequence identity to the amino acid sequence of SEQ ID NO:2.

Percent sequence identity is determined by conventional methods. See, for example, Altschul *et al.*, Bull. Math. Bio. 48:603 (1986), and Henikoff and Henikoff, 15 Proc. Natl. Acad. Sci. USA 89:10915 (1992). Briefly, two amino acid sequences are aligned to optimize the alignment scores using a gap opening penalty of 10, a gap extension penalty of 1, and the "BLOSUM62" scoring matrix of Henikoff and Henikoff (*ibid.*) as shown in Table 3 (amino acids are indicated by the standard one-letter codes).

Total number of identical matches

_____ x 100

[length of the longer sequence plus the
number of gaps introduced into the longer
sequence in order to align the two sequences]

25

Table 3

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
A	4																			
R	-1	5																		
N	-2	0	6																	
D	-2	-2	1	6																
C	0	-3	-3	-3	9															
Q	-1	1	0	0	-3	5														
E	-1	0	0	2	-4	2	5													
G	0	-2	0	-1	-3	-2	-2	6												
H	-2	0	1	-1	-3	0	0	-2	8											
I	-1	-3	-3	-3	-1	-3	-3	-4	-3	4										
L	-1	-2	-3	-4	-1	-2	-3	-4	-3	2	4									
K	-1	2	0	-1	-3	1	1	-2	-1	-3	-2	5								
M	-1	-1	-2	-3	-1	0	-2	-3	-2	1	2	-1	5							
F	-2	-3	-3	-2	-3	-3	-3	-1	0	0	-3	0	6							
P	-1	-2	-2	-1	-3	-1	-1	-2	-2	-3	-3	-1	-2	-4	7					
S	1	-1	1	0	-1	0	0	0	-1	-2	-2	0	-1	-2	-1	4				
T	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-2	-1	1	5				
W	-3	-3	-4	-4	-2	-2	-3	-2	-2	-3	-2	-3	-1	1	-4	-3	-2	11		
Y	-2	-2	-3	-2	-1	-2	-3	2	-1	-1	-2	-1	3	-3	-2	-2	2	7		
V	0	-3	-3	-3	-1	-2	-2	-3	-3	3	1	-2	1	-1	-2	0	-3	-1	4	

Those skilled in the art appreciate that there are many established algorithms available to align two amino acid sequences. The "FASTA" similarity search algorithm of Pearson and Lipman is a suitable protein alignment method for examining the level of identity shared by an amino acid sequence disclosed herein and the amino acid sequence of a putative variant zCTGF4. The FASTA algorithm is described by Pearson and Lipman, *Proc. Nat'l Acad. Sci. USA* 85:2444 (1988), and by Pearson, *Meth. Enzymol.* 183:63 (1990).

Briefly, FASTA first characterizes sequence similarity by identifying regions shared by the query sequence (e.g., SEQ ID NO:2) and a test sequence that have either the highest density of identities (if the ktup variable is 1) or pairs of identities (if ktup=2), without considering conservative amino acid substitutions, insertions, or deletions. The ten regions with the highest density of identities are then rescored by comparing the similarity of all paired amino acids using an amino acid substitution matrix, and the ends of the regions are "trimmed" to include only those residues that contribute to the highest score. If there are several regions with scores greater than the "cutoff" value (calculated by a predetermined formula based upon the length of the sequence and the ktup value), then the trimmed initial regions are examined to determine whether the regions can be joined to form an approximate alignment with gaps. Finally, the highest scoring regions of the two amino acid sequences are aligned using a modification of the Needleman-Wunsch-Sellers algorithm (Needleman and Wunsch, *J. Mol. Biol.* 48:444 (1970); Sellers, *SIAM J. Appl. Math.* 26:787 (1974)), which allows for amino acid insertions and deletions. Illustrative parameters for FASTA analysis are: ktup=1, gap opening penalty=10, gap extension penalty=1, and substitution matrix=BLOSUM62. These parameters can be introduced into a FASTA program by

modifying the scoring matrix file ("SMATRIX"), as explained in Appendix 2 of Pearson, *Meth. Enzymol.* 183:63 (1990).

FASTA can also be used to determine the sequence identity of nucleic acid molecules using a ratio as disclosed above. For nucleotide sequence comparisons, the ktup value can range between one to six, preferably from four to six.

Variant zCTGF4 polypeptides or polypeptides with substantially similar sequence identity are characterized as having one or more amino acid substitutions, deletions or additions. These changes are preferably of a minor nature, that is conservative amino acid substitutions (see Table 4) and other substitutions that do not significantly affect the folding or activity of the polypeptide; small deletions, typically of one to about 30 amino acids; and amino- or carboxyl-terminal extensions, such as an amino-terminal methionine residue, a small linker peptide of up to about 20-25 residues, or an affinity tag. The present invention thus includes polypeptides of from about 28 to 354 amino acid residues that comprise a sequence that is at least 70%, preferably at least 90%, and more preferably 95% or more identical to the corresponding region of SEQ ID NO:2. In particular, peptides and polypeptides corresponding to domains of the zCTGF4 molecules as shown in SEQ ID NO: 2 include the IBP domain (residues 59-102), the von Willebrand factor C domain (residues 114-179), the variable domain (residues 180-208) and the glycoconjugate binding domains (residues 209-252) are within the scope of the present invention. Polypeptides comprising affinity tags can further comprise a proteolytic cleavage site between the zCTGF4 polypeptide and the affinity tag. Preferred such sites include thrombin cleavage sites and factor Xa cleavage sites.

Table 4Conservative amino acid substitutions

5

	Basic:	arginine lysine histidine
10	Acidic:	glutamic acid aspartic acid
	Polar:	glutamine asparagine
	Hydrophobic:	leucine isoleucine
15		valine
	Aromatic:	phenylalanine tryptophan tyrosine
20	Small:	glycine alanine serine threonine methionine

25

The proteins of the present invention can also comprise non-naturally occurring amino acid residues. Non-naturally occurring amino acids include, without limitation, *trans*-3-methylproline, 2,4-methanoproline, *cis*-4-hydroxyproline, *trans*-4-hydroxyproline, *N*-methylglycine, *allo*-threonine, methylthreonine, hydroxyethylcysteine, hydroxyethylhomocysteine, nitroglutamine, homoglutamine, piperolic acid, thiazolidine carboxylic acid, dehydroproline, 3- and 4-methylproline, 3,3-dimethylproline, *tert*-leucine, norvaline, 2-azaphenylalanine, 3-azaphenylalanine, 4-azaphenylalanine, and 4-fluorophenylalanine. Several

30

35

methods are known in the art for incorporating non-naturally occurring amino acid residues into proteins. For example, an *in vitro* system can be employed wherein nonsense mutations are suppressed using chemically 5 aminoacylated suppressor tRNAs. Methods for synthesizing amino acids and aminoacylating tRNA are known in the art. Transcription and translation of plasmids containing nonsense mutations is typically carried out in a cell-free system comprising an *E. coli* S30 extract and commercially 10 available enzymes and other reagents. Proteins are purified by chromatography. See, for example, Robertson et al., *J. Am. Chem. Soc.* 113:2722 (1991), Ellman et al., *Methods Enzymol.* 202:301 (1991), Chung et al., *Science* 259:806 (1993), and Chung et al., *Proc. Nat'l Acad. Sci. USA* 90:10145 (1993).

In a second method, translation is carried out in *Xenopus* oocytes by microinjection of mutated mRNA and chemically aminoacylated suppressor tRNAs (Turcatti et al., *J. Biol. Chem.* 271:19991 (1996)). Within a third 20 method, *E. coli* cells are cultured in the absence of a natural amino acid that is to be replaced (e.g., phenylalanine) and in the presence of the desired non-naturally occurring amino acid(s) (e.g., 2-azaphenylalanine, 3-azaphenylalanine, 4-azaphenylalanine, 25 or 4-fluorophenylalanine). The non-naturally occurring amino acid is incorporated into the protein in place of its natural counterpart. See, Koide et al., *Biochem.* 33:7470 (1994). Naturally occurring amino acid residues can be converted to non-naturally occurring species by *in* 30 *vitro* chemical modification. Chemical modification can be combined with site-directed mutagenesis to further expand the range of substitutions (Wynn and Richards, *Protein Sci.* 2:395 (1993)).

A limited number of non-conservative amino 35 acids, amino acids that are not encoded by the genetic code, non-naturally occurring amino acids, and unnatural

amino acids may be substituted for zCTGF4 amino acid residues.

Multiple amino acid substitutions can be made and tested using known methods of mutagenesis and screening, such as those disclosed by Reidhaar-Olson and Sauer (*Science* 241:53 (1988)) or Bowie and Sauer (*Proc. Nat'l Acad. Sci. USA* 86:2152 (1989)). Briefly, these authors disclose methods for simultaneously randomizing two or more positions in a polypeptide, selecting for functional polypeptide, and then sequencing the mutagenized polypeptides to determine the spectrum of allowable substitutions at each position. Other methods that can be used include phage display (e.g., Lowman et al., *Biochem.* 30:10832 (1991), Ladner et al., U.S. Patent No. 5,223,409, Huse, international publication No. WO 92/06204, and region-directed mutagenesis (Derbyshire et al., *Gene* 46:145 (1986), and Ner et al., *DNA* 7:127, (1988)).

Variants of the disclosed zCTGF4 nucleotide and polypeptide sequences can also be generated through DNA shuffling as disclosed by Stemmer, *Nature* 370:389 (1994), Stemmer, *Proc. Nat'l Acad. Sci. USA* 91:10747 (1994), and international publication No. WO 97/20078. Briefly, variant DNA molecules are generated by *in vitro* homologous recombination by random fragmentation of a parent DNA followed by reassembly using PCR, resulting in randomly introduced point mutations. This technique can be modified by using a family of parent DNA molecules, such as allelic variants or DNA molecules from different species, to introduce additional variability into the process. Selection or screening for the desired activity, followed by additional iterations of mutagenesis and assay provides for rapid "evolution" of sequences by selecting for desirable mutations while simultaneously selecting against detrimental changes.

Mutagenesis methods as disclosed herein can be combined with high-throughput, automated screening methods to detect activity of cloned, mutagenized polypeptides in host cells. Mutagenized DNA molecules that encode 5 biologically active polypeptides, or polypeptides that bind with anti-zCTGF4 antibodies, can be recovered from the host cells and rapidly sequenced using modern equipment. These methods allow the rapid determination of 10 the importance of individual amino acid residues in a polypeptide of interest, and can be applied to polypeptides of unknown structure.

Essential amino acids in the polypeptides of the present invention can be identified according to procedures known in the art, such as site-directed 15 mutagenesis or alanine-scanning mutagenesis (Cunningham and Wells, *Science* 244:1081 (1989), Bass et al., *Proc. Nat'l Acad. Sci. USA* 88:4498 (1991), Coombs and Corey, "Site-Directed Mutagenesis and Protein Engineering," in *Proteins: Analysis and Design*, Angeletti (ed.), pages 259- 20 311 (Academic Press, Inc. 1998)). In the latter technique, single alanine mutations are introduced at every residue in the molecule, and the resultant mutant molecules are tested for biological activity as disclosed below to identify amino acid residues that are critical to 25 the activity of the molecule. See also, Hilton et al., *J. Biol. Chem.* 271:4699 (1996). The identities of essential amino acids can also be inferred from analysis of homologies with zCTGF4.

The location of zCTGF4 receptor binding domains 30 can be identified by physical analysis of structure, as determined by such techniques as nuclear magnetic resonance, crystallography, electron diffraction or photoaffinity labeling, in conjunction with mutation of putative contact site amino acids. See, for example, de 35 Vos et al., *Science* 255:306 (1992), Smith et al., *J. Mol. Biol.* 224:899 (1992), and Wlodaver et al., *FEBS Lett.*

309:59 (1992). Moreover, zCTGF4 labeled with biotin or FITC can be used for expression cloning of zCTGF4 receptors.

The present invention also includes "functional fragments" of zCTGF4 polypeptides and nucleic acid molecules encoding such functional fragments. As previously described herein, zCTGF4 is characterized by a multi-domain structure comprising a IBP domain (amino acid residue 59 (Pro) to 102 (Tyr) of SEQ ID NO: 2) that has been suggested as an insulin growth factor binding domain, a von Willebrand factor c domain (amino acid residue 114 (Cys) to 179 (Cys) of SEQ ID NO: 2), a variable domain (amino acid 180 (Ser) to 208 (Lys), and a sulfated glycoconjugate binding motif domain (amino acid residue 209 (Cys) to 252 (Cys) of SEQ ID NO: 2). Thus, the present invention further provides fusion proteins encompassing (a) polypeptide molecules comprising one or more of the domains described above, and (b) biologically active fragments comprising portions of one or more of the domains. The other polypeptide may be another domains from another CTGF, a non-native and/or an unrelated secretory signal peptide to facilitate secretion of the fusion protein.

Routine deletion analyses of nucleic acid molecules can be performed to obtain functional fragments of a nucleic acid molecule that encodes an zCTGF4 polypeptide. As an illustration, DNA molecules having the nucleotide sequence of SEQ ID NO:1 can be digested with *Bal31* nuclease to obtain a series of nested deletions. The fragments are then inserted into expression vectors in proper reading frame, and the expressed polypeptides are isolated and tested for zCTGF4, or for the ability to bind anti-zCTGF4 antibodies. One alternative to exonuclease digestion is to use oligonucleotide-directed mutagenesis to introduce deletions or stop codons to specify production of a desired fragment. Alternatively,

particular fragments of an zCTGF4 gene can be synthesized using the polymerase chain reaction.

Standard methods for identifying functional domains are well-known to those of skill in the art. For example, studies on the truncation at either or both termini of interferons have been summarized by Horisberger and Di Marco, *Pharmac. Ther.* 66:507 (1995). Moreover, standard techniques for functional analysis of proteins are described by, for example, Treuter et al., *Molec. Gen. Genet.* 240:113 (1993), Content et al., "Expression and preliminary deletion analysis of the 42 kDa 2-5A synthetase induced by human interferon," in *Biological Interferon Systems, Proceedings of ISIR-TNO Meeting on Interferon Systems*, Cantell (ed.), pages 65-72 (Nijhoff 1987), Herschman, "The EGF Receptor," in *Control of Animal Cell Proliferation*, Vol. 1, Boynton et al., (eds.) pages 169-199 (Academic Press 1985), Coumailleau et al., *J. Biol. Chem.* 270:29270 (1995); Fukunaga et al., *J. Biol. Chem.* 270:25291 (1995); Yamaguchi et al., *Biochem. Pharmacol.* 50:1295 (1995), and Meisel et al., *Plant Molec. Biol.* 30:1 (1996).

The present invention also contemplates functional fragments of an zCTGF4 gene that has amino acid changes, compared with the amino acid sequence of SEQ ID NO:2. A variant zCTGF4 gene can be identified on the basis of structure by determining the level of identity with nucleotide and amino acid sequences of SEQ ID NOS:1 and 2, as discussed above. An alternative approach to identifying a variant gene on the basis of structure is to determine whether a nucleic acid molecule encoding a potential variant zCTGF4 gene can hybridize to a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1, as discussed above.

The present invention also provides polypeptide fragments or peptides comprising an epitope-bearing portion of an zCTGF4 polypeptide described herein. Such

fragments or peptides may comprise an "immunogenic epitope," which is a part of a protein that elicits an antibody response when the entire protein is used as an immunogen. Immunogenic epitope-bearing peptides can be
5 identified using standard methods (see, for example, Geysen et al., *Proc. Nat'l Acad. Sci. USA* 81:3998 (1983)).

In contrast, polypeptide fragments or peptides may comprise an "antigenic epitope," which is a region of a protein molecule to which an antibody can specifically
10 bind. Certain epitopes consist of a linear or contiguous stretch of amino acids, and the antigenicity of such an epitope is not disrupted by denaturing agents. It is known in the art that relatively short synthetic peptides that can mimic epitopes of a protein can be used to stimulate
15 the production of antibodies against the protein (see, for example, Sutcliffe et al., *Science* 219:660 (1983)). Accordingly, antigenic epitope-bearing peptides and polypeptides of the present invention are useful to raise
20 antibodies that bind with the polypeptides described herein.

Antigenic epitope-bearing peptides and polypeptides preferably contain at least four to ten amino acids, at least ten to fifteen amino acids, or about 15 to about 30 amino acids of SEQ ID NO:2. Such epitope-bearing
25 peptides and polypeptides can be produced by fragmenting an zCTGF4 polypeptide, or by chemical peptide synthesis, as described herein. Moreover, epitopes can be selected by phage display of random peptide libraries (see, for example, Lane and Stephen, *Curr. Opin. Immunol.* 5:268
30 (1993), and Cortese et al., *Curr. Opin. Biotechnol.* 7:616 (1996)). Standard methods for identifying epitopes and producing antibodies from small peptides that comprise an epitope are described, for example, by Mole, "Epitope Mapping," in *Methods in Molecular Biology*, Vol. 10, Manson
35 (ed.), pages 105-116 (The Humana Press, Inc. 1992), Price, "Production and Characterization of Synthetic Peptide-

Derived Antibodies," in *Monoclonal Antibodies: Production, Engineering, and Clinical Application*, Ritter and Ladyman (eds.), pages 60-84 (Cambridge University Press 1995), and Coligan *et al.* (eds.), *Current Protocols in Immunology*, 5 pages 9.3.1 - 9.3.5 and pages 9.4.1 - 9.4.11 (John Wiley & Sons 1997).

Regardless of the particular nucleotide sequence of a variant zCTGF4 gene, the gene encodes a polypeptide that is characterized by its proliferative or 10 differentiating activity, or ability to induce specialized cell functions, or by the ability to bind specifically to an anti-zCTGF4 antibody. More specifically, variant zCTGF4 genes encode polypeptides which exhibit at least 50% and preferably, greater than 70, 80 or 90%, of the 15 activity of polypeptide encoded by the human zCTGF4 gene described herein.

For any zCTGF4 polypeptide, including variants and fusion proteins, one of ordinary skill in the art can readily generate a fully degenerate polynucleotide 20 sequence encoding that variant using the information set forth in Tables 1 and 2 above.

The present invention further provides a variety of other polypeptide fusions (and related multimeric proteins comprising one or more polypeptide fusions). For 25 example, a zCTGF4 polypeptide can be prepared as a fusion to a dimerizing protein as disclosed in U.S. Patents Nos. 5,155,027 and 5,567,584. Preferred dimerizing proteins in this regard include immunoglobulin constant region domains. Immunoglobulin- zCTGF4 polypeptide fusions can 30 be expressed in genetically engineered cells (to produce a variety of multimeric zCTGF4 analogs). Auxiliary domains can be fused to zCTGF4 polypeptides to target them to specific cells, tissues, or macromolecules (e.g., collagen). For example, a zCTGF4 polypeptide or protein 35 could be targeted to a predetermined cell type by fusing a zCTGF4 polypeptide to a ligand that specifically binds to a receptor on the surface of the target cell. In this

way, polypeptides and proteins can be targeted for therapeutic or diagnostic purposes. A zCTGF4 polypeptide can be fused to two or more moieties, such as an affinity tag for purification and a targeting domain. Polypeptide fusions can also comprise one or more cleavage sites, particularly between domains. See, Tuan et al., Connective Tissue Research 34:1-9, 1996.

A Hopp/Woods hydrophilicity profile of the zCTGF4 protein sequence as shown in SEQ ID NO:2 can be generated (Hopp et al., Proc. Natl. Acad. Sci. 78:3824-3828, 1981; Hopp, J. Immun. Meth. 88:1-18, 1986 and Triquier et al., Protein Engineering 11:153-169, 1998). The profile is based on a sliding six-residue window. Buried G, S, and T residues and exposed H, Y, and W residues were ignored. Hydrophilicity can be used to determine regions that have the most antigenic potential. For example, in zCTGF4, hydrophilic regions include amino acid residues 239-244 of SEQ ID NO: 2, amino acid residues 105-110 of SEQ ID NO: 2, amino acid residues 172-177 of SEQ ID NO: 2, amino acid residues 238-243 of SEQ ID NO: 2, and amino acid residues 171-176 of SEQ ID NO: 2.

Using the methods discussed herein, one of ordinary skill in the art can identify and/or prepare a variety of polypeptides that are have substantially similar sequence identity to residues 1 or 24 to 354 of SEQ ID NO: 2, or functional fragments and fusions thereof, and retain the properties of the wild-type protein such as the ability to stimulate proliferation, differentiation or induce specialized cell function.

The polypeptides of the present invention, including full-length proteins, fragments thereof and fusion proteins, can be produced in genetically engineered host cells according to conventional techniques. Suitable host cells are those cell types that can be transformed or transfected with exogenous DNA and grown in culture, and include bacteria, fungal cells, and cultured higher eukaryotic cells. Eukaryotic cells, particularly cultured

cells of multicellular organisms, are preferred. Techniques for manipulating cloned DNA molecules and introducing exogenous DNA into a variety of host cells are disclosed by Sambrook et al., Molecular Cloning: A 5 Laboratory Manual, 2nd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989, and Ausubel et al., eds., Current Protocols in Molecular Biology, John Wiley and Sons, Inc., NY, 1987.

In general, a DNA sequence encoding a zCTGF4 10 polypeptide is operably linked to other genetic elements required for its expression, generally including a transcription promoter and terminator, within an expression vector. The vector will also commonly contain one or more selectable markers and one or more origins of 15 replication, although those skilled in the art will recognize that within certain systems selectable markers may be provided on separate vectors, and replication of the exogenous DNA may be provided by integration into the host cell genome. Selection of promoters, terminators, 20 selectable markers, vectors and other elements is a matter of routine design within the level of ordinary skill in the art. Many such elements are described in the literature and are available through commercial suppliers.

To direct a zCTGF4 polypeptide into the 25 secretory pathway of a host cell, a secretory signal sequence (also known as a leader sequence, prepro sequence or pre sequence) is provided in the expression vector. The secretory signal sequence may be that of the zCTGF4 30 polypeptide, or may be derived from another secreted protein (e.g., t-PA) or synthesized de novo. The secretory signal sequence is operably linked to the zCTGF4 DNA sequence, i.e., the two sequences are joined in the correct reading frame and positioned to direct the newly 35 synthesized polypeptide into the secretory pathway of the host cell. Secretory signal sequences are commonly positioned 5' to the DNA sequence encoding the polypeptide

of interest, although certain secretory signal sequences may be positioned elsewhere in the DNA sequence of interest (see, e.g., Welch et al., U.S. Patent No. 5,037,743; Holland et al., U.S. Patent No. 5,143,830).

5 Alternatively, the secretory signal sequence contained in the polypeptides of the present invention is used to direct other polypeptides into the secretory pathway. The present invention provides for such fusion polypeptides. A signal fusion polypeptide can be made
10 wherein a secretory signal sequence derived from amino acid residue 1 to 23 of SEQ ID NO:2 is operably linked to a DNA sequence encoding another polypeptide using methods known in the art and disclosed herein. The secretory signal sequence contained in the fusion polypeptides of
15 the present invention is preferably fused amino-terminally to an additional peptide to direct the additional peptide into the secretory pathway. Such constructs have numerous applications known in the art. For example, these novel secretory signal sequence fusion constructs can direct the
20 secretion of an active component of a normally non-secreted protein. Such fusions may be used *in vivo* or *in vitro* to direct peptides through the secretory pathway.

Cultured mammalian cells are suitable hosts within the present invention. Methods for introducing exogenous DNA into mammalian host cells include calcium phosphate-mediated transfection (Wigler et al., Cell 14:725, 1978; Corsaro and Pearson, Somatic Cell Genetics 7:603, 1981; Graham and Van der Eb, Virology 52:456, 1973), electroporation (Neumann et al., EMBO J. 1:841-845, 1982), DEAE-dextran mediated transfection (Ausubel et al., ibid.), and liposome-mediated transfection (Hawley-Nelson et al., Focus 15:73, 1993; Ciccarone et al., Focus 15:80, 1993), and viral vectors (Miller and Rosman, BioTechniques 7:980-90, 1989; Wang and Finer, Nature Med. 2:714-6, 1996). The production of recombinant polypeptides in cultured mammalian cells is disclosed, for example, by

Levinson et al., U.S. Patent No. 4,713,339; Hagen et al., U.S. Patent No. 4,784,950; Palmiter et al., U.S. Patent No. 4,579,821; and Ringold, U.S. Patent No. 4,656,134. Suitable cultured mammalian cells include the COS-1 (ATCC No. CRL 1650), COS-7 (ATCC No. CRL 1651), BHK (ATCC No. CRL 1632), BHK 570 (ATCC No. CRL 10314), 293 (ATCC No. CRL 1573; Graham et al., J. Gen. Virol. 36:59-72, 1977) and Chinese hamster ovary (e.g. CHO-K1; ATCC No. CCL 61) cell lines. Additional suitable cell lines are known in the art and available from public depositories such as the American Type Culture Collection, Manassas, VA. In general, strong transcription promoters are preferred, such as promoters from SV-40 or cytomegalovirus. See, e.g., U.S. Patent No. 4,956,288. Other suitable promoters include those from metallothionein genes (U.S. Patent Nos. 4,579,821 and 4,601,978) and the adenovirus major late promoter.

Drug selection is generally used to select for cultured mammalian cells into which foreign DNA has been inserted. Such cells are commonly referred to as "transfectants". Cells that have been cultured in the presence of the selective agent and are able to pass the gene of interest to their progeny are referred to as "stable transfectants." A preferred selectable marker is a gene encoding resistance to the antibiotic neomycin. Selection is carried out in the presence of a neomycin-type drug, such as G-418 or the like. Selection systems can also be used to increase the expression level of the gene of interest, a process referred to as "amplification." Amplification is carried out by culturing transfectants in the presence of a low level of the selective agent and then increasing the amount of selective agent to select for cells that produce high levels of the products of the introduced genes. A preferred amplifiable selectable marker is dihydrofolate reductase, which confers resistance to methotrexate.

Other drug resistance genes (e.g. hygromycin resistance, multi-drug resistance, puromycin acetyltransferase) can also be used. Alternative markers that introduce an altered phenotype, such as green fluorescent protein, or .5 cell surface proteins such as CD4, CD8, Class I MHC, placental alkaline phosphatase may be used to sort transfected cells from untransfected cells by such means as FACS sorting or magnetic bead separation technology.

Other higher eukaryotic cells can also be used 10 as hosts, including plant cells, insect cells and avian cells. The use of *Agrobacterium rhizogenes* as a vector for expressing genes in plant cells has been reviewed by Sinkar et al., J. Biosci. (Bangalore) 11:47-58, 1987. Transformation of insect cells and production of foreign 15 polypeptides therein is disclosed by Guarino et al., U.S. Patent No. 5,162,222 and WIPO publication WO 94/06463. Insect cells can be infected with recombinant baculovirus, commonly derived from *Autographa californica nuclear polyhedrosis virus* (AcNPV). DNA encoding the zCTGF4 20 polypeptide is inserted into the baculoviral genome in place of the AcNPV polyhedrin gene coding sequence by one of two methods. The first is the traditional method of homologous DNA recombination between wild-type AcNPV and a transfer vector containing the zCTGF4 flanked by AcNPV 25 sequences. Suitable insect cells, e.g. SF9 cells, are infected with wild-type AcNPV and transfected with a transfer vector comprising a zCTGF4 polynucleotide operably linked to an AcNPV polyhedrin gene promoter, terminator, and flanking sequences. See, King, L.A. and 30 Possee, R.D., The Baculovirus Expression System: A Laboratory Guide, London, Chapman & Hall; O'Reilly, D.R. et al., Baculovirus Expression Vectors: A Laboratory Manual, New York, Oxford University Press., 1994; and, Richardson, C. D., Ed., Baculovirus Expression Protocols. 35 Methods in Molecular Biology, Totowa, NJ, Humana Press,

1995. Natural recombination within an insect cell will result in a recombinant baculovirus which contains zCTGF4 driven by the polyhedrin promoter. Recombinant viral stocks are made by methods commonly used in the art.

5 The second method of making recombinant baculovirus utilizes a transposon-based system described by Luckow (Luckow, V.A, et al., J Virol 67:4566-79, 1993). This system is sold in the Bac-to-Bac kit (Life Technologies, Rockville, MD). This system utilizes a
10 transfer vector, pFastBacTM (Life Technologies) containing a Tn7 transposon to move the DNA encoding the zCTGF4 polypeptide into a baculovirus genome maintained in E. coli as a large plasmid called a "bacmid." The pFastBacTM transfer vector utilizes the AcNPV polyhedrin
15 promoter to drive the expression of the gene of interest, in this case zCTGF4. However, pFastBacTM can be modified to a considerable degree. The polyhedrin promoter can be removed and substituted with the baculovirus basic protein promoter (also known as Pcor, p6.9 or MP promoter) which
20 is expressed earlier in the baculovirus infection, and has been shown to be advantageous for expressing secreted proteins. See, Hill-Perkins, M.S. and Possee, R.D., J. Gen. Virol. 71:971-6, 1990; Bonning, B.C. et al., J. Gen. Virol. 75:1551-6, 1994; and, Chazenbalk, G.D., and
25 Rapoport, B., J. Biol. Chem. 270:1543-9, 1995. In such transfer vector constructs, a short or long version of the basic protein promoter can be used. Moreover, transfer vectors can be constructed which replace the native zCTGF4 secretory signal sequences with secretory signal sequences
30 derived from insect proteins. For example, a secretory signal sequence from Ecdysteroid Glucosyltransferase (EGT), honey bee Melittin (Invitrogen, Carlsbad, CA), or baculovirus gp67 (PharMingen, San Diego, CA) can be used in constructs to replace the native zCTGF4 secretory signal sequence. In addition, transfer vectors can include an in-frame fusion with DNA encoding an epitope
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tag at the C- or N-terminus of the expressed zCTGF4 polypeptide, for example, a Glu-Glu epitope tag (Grussenmeyer, T. et al., Proc. Natl. Acad. Sci. 82:7952-4, 1985). Using a technique known in the art, a transfer vector containing zCTGF4 is transformed into E. Coli, and screened for bacmids which contain an interrupted lacZ gene indicative of recombinant baculovirus. The bacmid DNA containing the recombinant baculovirus genome is isolated, using common techniques, and used to transfect *Spodoptera frugiperda* cells, e.g. Sf9 cells. Recombinant virus that expresses zCTGF4 is subsequently produced. Recombinant viral stocks are made by methods commonly used the art.

The recombinant virus is used to infect host cells, typically a cell line derived from the fall armyworm, *Spodoptera frugiperda*. See, in general, Glick and Pasternak, Molecular Biotechnology: Principles and Applications of Recombinant DNA, ASM Press, Washington, D.C., 1994. Another suitable cell line is the High FiveO™ cell line (Invitrogen) derived from *Trichoplusia ni* (U.S. Patent No. 5,300,435). Commercially available serum-free media are used to grow and maintain the cells. Suitable media are Sf900 II™ (Life Technologies) or ESF 921™ (Expression Systems) for the Sf9 cells; and Express 5 cell0405™ (JRH Biosciences, Lenexa, KS) or Express FiveO™ (Life Technologies) for the *T. ni* cells. The cells are grown up from an inoculation density of approximately 2-5 $\times 10^5$ cells to a density of 1-2 $\times 10^6$ cells at which time a recombinant viral stock is added at a multiplicity of infection (MOI) of 0.1 to 10, more typically near 3. The recombinant virus-infected cells typically produce the recombinant zCTGF4 polypeptide at 12-72 hours post-infection and secrete it with varying efficiency into the medium. The culture is usually harvested 48 hours post-infection. Centrifugation is used to separate the cells

from the medium (supernatant). The supernatant containing the zCTGF4 polypeptide is filtered through micropore filters, usually 0.45 μm pore size. Procedures used are generally described in available laboratory manuals (King,
5 L. A. and Possee, R.D., ibid.; O'Reilly, D.R. et al., ibid.; Richardson, C. D., ibid.). Subsequent purification of the zCTGF4 polypeptide from the supernatant can be achieved using methods described herein.

Fungal cells, including yeast cells, can also be
10 used within the present invention. Yeast species of particular interest in this regard include *Saccharomyces cerevisiae*, *Pichia pastoris*, and *Pichia methanolica*. Methods for transforming *S. cerevisiae* cells with exogenous DNA and producing recombinant polypeptides
15 therefrom are disclosed by, for example, Kawasaki, U.S. Patent No. 4,599,311; Kawasaki et al., U.S. Patent No. 4,931,373; Brake, U.S. Patent No. 4,870,008; Welch et al., U.S. Patent No. 5,037,743; and Murray et al., U.S. Patent No. 4,845,075. Transformed cells are selected by
20 phenotype determined by the selectable marker, commonly drug resistance or the ability to grow in the absence of a particular nutrient (e.g., leucine). A preferred vector system for use in *Saccharomyces cerevisiae* is the POT1 vector system disclosed by Kawasaki et al. (U.S. Patent
25 No. 4,931,373), which allows transformed cells to be selected by growth in glucose-containing media. Suitable promoters and terminators for use in yeast include those from glycolytic enzyme genes (see, e.g., Kawasaki, U.S. Patent No. 4,599,311; Kingsman et al., U.S. Patent No.
30 4,615,974; and Bitter, U.S. Patent No. 4,977,092) and alcohol dehydrogenase genes. See also U.S. Patents Nos. 4,990,446; 5,063,154; 5,139,936 and 4,661,454. Transformation systems for other yeasts, including
35 *Hansenula polymorpha*, *Schizosaccharomyces pombe*, *Kluyveromyces lactis*, *Kluyveromyces fragilis*, *Ustilago*

maydis, *Pichia pastoris*, *Pichia methanolica*, *Pichia guillermondii* and *Candida maltosa* are known in the art. See, for example, Gleeson et al., J. Gen. Microbiol. 132:3459-65, 1986 and Clegg, U.S. Patent No. 4,882,279.

5 *Aspergillus* cells may be utilized according to the methods of McKnight et al., U.S. Patent No. 4,935,349. Methods for transforming *Acremonium chrysogenum* are disclosed by Sumino et al., U.S. Patent No. 5,162,228. Methods for transforming *Neurospora* are disclosed by Lambowitz, U.S.

10 Patent No. 4,486,533.

The use of *Pichia methanolica* as host for the production of recombinant proteins is disclosed in WIPO Publications WO 97/17450, WO 97/17451, WO 98/02536, and WO 98/02565. DNA molecules for use in transforming *P. methanolica* will commonly be prepared as double-stranded, circular plasmids, which are preferably linearized prior to transformation. For polypeptide production in *P. methanolica*, it is preferred that the promoter and terminator in the plasmid be that of a *P. methanolica* gene, such as a *P. methanolica* alcohol utilization gene (AUG1 or AUG2). Other useful promoters include those of the dihydroxyacetone synthase (DHAS), formate dehydrogenase (FMD), and catalase (CAT) genes. To facilitate integration of the DNA into the host chromosome, it is preferred to have the entire expression segment of the plasmid flanked at both ends by host DNA sequences. A preferred selectable marker for use in *Pichia methanolica* is a *P. methanolica* ADE2 gene, which encodes phosphoribosyl-5-aminoimidazole carboxylase (AIRC; EC 4.1.1.21), which allows ade2 host cells to grow in the absence of adenine. For large-scale, industrial processes where it is desirable to minimize the use of methanol, it is preferred to use host cells in which both methanol utilization genes (AUG1 and AUG2) are deleted. For

production of secreted proteins, host cells deficient in vacuolar protease genes (*PEP4* and *PRB1*) are preferred. Electroporation is used to facilitate the introduction of a plasmid containing DNA encoding a polypeptide of interest into *P. methanolica* cells. It is preferred to transform *P. methanolica* cells by electroporation using an exponentially decaying, pulsed electric field having a field strength of from 2.5 to 4.5 kV/cm, preferably about 3.75 kV/cm, and a time constant (τ) of from 1 to 40 milliseconds, most preferably about 20 milliseconds.

Prokaryotic host cells, including strains of the bacteria *Escherichia coli*, *Bacillus* and other genera are also useful host cells within the present invention. Techniques for transforming these hosts and expressing foreign DNA sequences cloned therein are well known in the art (see, e.g., Sambrook et al., *ibid.*). When expressing a zCTGF4 polypeptide in bacteria such as *E. coli*, the polypeptide may be retained in the cytoplasm, typically as insoluble granules, or may be directed to the periplasmic space by a bacterial secretion sequence. In the former case, the cells are lysed, and the granules are recovered and denatured using, for example, guanidine isothiocyanate or urea. The denatured polypeptide can then be refolded and dimerized by diluting the denaturant, such as by dialysis against a solution of urea and a combination of reduced and oxidized glutathione, followed by dialysis against a buffered saline solution. In the latter case, the polypeptide can be recovered from the periplasmic space in a soluble and functional form by disrupting the cells (by, for example, sonication or osmotic shock) to release the contents of the periplasmic space and recovering the protein, thereby obviating the need for denaturation and refolding.

Transformed or transfected host cells are cultured according to conventional procedures in a culture medium containing nutrients and other components required

71 (Toso Haas) and the like. Suitable solid supports include glass beads, silica-based resins, cellulosic resins, agarose beads, cross-linked agarose beads, polystyrene beads, cross-linked polyacrylamide resins and 5 the like that are insoluble under the conditions in which they are to be used. These supports may be modified with reactive groups that allow attachment of proteins by amino groups, carboxyl groups, sulfhydryl groups, hydroxyl groups and/or carbohydrate moieties. Examples of coupling 10 chemistries include cyanogen bromide activation, N-hydroxysuccinimide activation, epoxide activation, sulfhydryl activation, hydrazide activation, and carboxyl and amino derivatives for carbodiimide coupling chemistries. These and other solid media are well known 15 and widely used in the art, and are available from commercial suppliers. Methods for binding receptor polypeptides to support media are well known in the art. Selection of a particular method is a matter of routine design and is determined in part by the properties of the 20 chosen support. See, for example, Affinity Chromatography: Principles & Methods, Pharmacia LKB Biotechnology, Uppsala, Sweden, 1988.

The polypeptides of the present invention can be isolated by exploitation of size, charge and 25 hydrophobicity. For example, immobilized metal ion adsorption (IMAC) chromatography can be used to purify histidine-rich proteins (E. Sulkowski, Trends in Biochem. 3:1-7, 1985). Other methods of purification include 30 purification of glycosylated proteins by lectin affinity chromatography and ion exchange chromatography (Methods in Enzymol., Vol. 182, "Guide to Protein Purification", M. Deutscher, (ed.), Acad. Press, San Diego, 1990, pp.529-39). Within additional embodiments of the invention, a 35 fusion of the polypeptide of interest and an affinity tag (e.g., maltose-binding protein, an immunoglobulin domain) may be constructed to facilitate purification. ZCTGF4 has a domain homologous to the heparin binding domain

described previously for CTGF, and exploitation of this property may be useful for purification of zCTGF4. For a review, see, Burgess et al., Ann. Rev. of Biochem. 58:575-606, 1989. Members of the FGF family, which also have a heparin binding domain can be purified to apparent homogeneity by heparin-Sepharose affinity chromatography (Gospodarowicz et al., Proc. Natl. Acad. Sci. 81:6963-6967, 1984) and eluted using linear step gradients of NaCl (Ron et al., J. Biol. Chem. 268(4):2984-2988, 1993; 10 Chromatography: Principles & Methods, pp. 77-80, Pharmacia LKB Biotechnology, Uppsala, Sweden, 1993; in "Immobilized Affinity Ligand Techniques", Hermanson et al., eds., pp. 165-167, Academic Press, San Diego, 1992; Kjellen et al., Ann. Rev. Biochem. Ann. Rev. Biochem. 15 60:443-474, 1991; and Ke et al., Protein Expr. Purif. 3(6):497-507, 1992.)

Protein refolding (and optionally reoxidation) procedures may be advantageously used. It is preferred to purify the protein to >80% purity, more preferably to >90% 20 purity, even more preferably >95%, and particularly preferred is a pharmaceutically pure state, that is greater than 99.9% pure with respect to contaminating macromolecules, particularly other proteins and nucleic acids, and free of infectious and pyrogenic agents. 25 Preferably, a purified protein is substantially free of other proteins, particularly other proteins of animal origin.

ZCTGF4 polypeptides or fragments thereof may also be prepared through chemical synthesis (Merrifield, 30 J. Am. Chem. Soc. 85:2149, 1963). ZCTGF4 polypeptides may be monomers or multimers; glycosylated or non-glycosylated; pegylated or non-pegylated; and may or may not include an initial methionine amino acid residue.

The activity of molecules of the present 35 invention can be measured using a variety of assays that measure cell proliferation, differentiation, chemotaxis or induction of specialized cell functions. Of particular

interest are changes in proliferation or differentiation of endothelial cells, particularly endothelial cells isolated from testis, trachea, bone marrow or kidney tissue. Proliferation and differentiation can be measured 5 using *in vitro* using cultured cells or *in vivo* by administering molecules of the claimed invention to the appropriate animal model. Assays measuring cell proliferation or differentiation are well known in the art. For example, assays measuring proliferation include 10 such assays as chemosensitivity to neutral red dye (Cavanaugh et al., Investigational New Drugs 8:347-354, 1990, incorporated herein by reference), incorporation of radiolabelled nucleotides (Cook et al., Analytical Biochem. 179:1-7, 1989, incorporated herein by reference), 15 incorporation of 5-bromo-2'-deoxyuridine (BrdU) in the DNA of proliferating cells (Porstmann et al., J. Immunol. Methods 82:169-179, 1985, incorporated herein by reference), and use of tetrazolium salts (Mosmann, J. Immunol. Methods 65:55-63, 1983; Alley et al., Cancer Res. 48:589-601, 1988; Marshall et al., Growth Reg. 5:69-84, 1995; and Scudiero et al., Cancer Res. 48:4827-4833, 1988; all incorporated herein by reference). Assays measuring 20 differentiation include, for example, measuring cell-surface markers associated with stage-specific expression of a tissue, enzymatic activity, functional activity or morphological changes (Watt, FASEB, 5:281-284, 1991; Francis, Differentiation 57:63-75, 1994; Raes, Adv. Anim. Cell Biol. Technol. Bioprocesses, 161-171, 1989; all incorporated herein by reference).

30 Examples of assays measuring induction of specialized cell functions include: extracellular matrix protein mRNA induction assays (Frazier et al., J. Invest. Dermatol. 107:404-411, 1996); ^{35}S methionine pulse-chase assays measuring stimulation of matrix protein synthesis 35 (Frazier et al., *ibid.*, 1996); subcutaneous administration of growth factors to mice (Roberts et al., Proc. Natl.

Acad. Sci. USA 83:4167-4171, 1986); and *in situ* hybridization to measure changes in mRNA expression (Fava et al., Blood 76:1946-1955, 1990).

An exemplary *in vivo* assay is when mammalian 5 transfected (or co-transfected) expression host cells may be embedded in an alginate environment and injected (implanted) into recipient animals. Alginate-poly-L-lysine microencapsulation, permselective membrane encapsulation and diffusion chambers have been described 10 as a means to entrap transfected mammalian cells or primary mammalian cells. These types of non-immunogenic "encapsulations" or microenvironments permit the transfer of nutrients into the microenvironment, and also permit the diffusion of proteins and other macromolecules 15 secreted or released by the captured cells across the environmental barrier to the recipient animal. Most importantly, the capsules or microenvironments mask and shield the foreign, embedded cells from the recipient animal's immune response. Such microenvironments can 20 extend the life of the injected cells from a few hours or days (naked cells) to several weeks (embedded cells).

An *in vivo* approach for assaying proteins of the present invention involves viral delivery systems. Exemplary viruses for this purpose include adenovirus, 25 herpesvirus, retroviruses, vaccinia virus, and adeno-associated virus (AAV). Adenovirus, a double-stranded DNA virus, is currently the best studied gene transfer vector for delivery of heterologous nucleic acid (for review, see Becker et al., Meth. Cell Biol. 43:161-89, 1994; and 30 Douglas and Curiel, Science & Medicine 4:44-53, 1997). The adenovirus system offers several advantages: (i) adenovirus can accommodate relatively large DNA inserts; (ii) can be grown to high-titer; (iii) infect a broad range of mammalian cell types; and (iv) can be used with 35 many different promoters including ubiquitous, tissue

specific, and regulatable promoters. Also, because adenoviruses are stable in the bloodstream, they can be administered by intravenous injection.

Using adenovirus vectors where portions of the adenovirus genome are deleted, inserts are incorporated into the viral DNA by direct ligation or by homologous recombination with a co-transfected plasmid. In an exemplary system, the essential E1 gene has been deleted from the viral vector, and the virus will not replicate unless the E1 gene is provided by the host cell (the human 293 cell line is exemplary). When intravenously administered to intact animals, adenovirus primarily targets the liver. If the adenoviral delivery system has an E1 gene deletion, the virus cannot replicate in the host cells. However, the host's tissue (e.g., liver) will express and process (and, if a secretory signal sequence is present, secrete) the heterologous protein. Secreted proteins will enter the circulation in the highly vascularized liver, and effects on the infected animal can be determined.

Moreover, adenoviral vectors containing various deletions of viral genes can be used in an attempt to reduce or eliminate immune responses to the vector. Such adenoviruses are E1 deleted, and in addition contain deletions of E2A or E4 (Lusky, M. et al., J. Virol. 72:2022-2032, 1998; Raper, S.E. et al., Human Gene Therapy 9:671-679, 1998). In addition, deletion of E2b is reported to reduce immune responses (Amalfitano, A. et al., J. Virol. 72:926-933, 1998). Moreover, by deleting the entire adenovirus genome, very large inserts of heterologous DNA can be accommodated. Generation of so called "gutless" adenoviruses where all viral genes are deleted are particularly advantageous for insertion of

large inserts of heterologous DNA. For review, see Yeh, P. and Perricaudet, M., FASEB J. 11:615-623, 1997.

The adenovirus system can also be used for protein production *in vitro*. By culturing adenovirus-infected non-293 cells under conditions where the cells are not rapidly dividing, the cells can produce proteins for extended periods of time. For instance, BHK cells are grown to confluence in cell factories, then exposed to the adenoviral vector encoding the secreted protein of interest. The cells are then grown under serum-free conditions, which allows infected cells to survive for several weeks without significant cell division. Alternatively, adenovirus vector infected 293 cells can be grown as adherent cells or in suspension culture at relatively high cell density to produce significant amounts of protein (See Garnier et al., Cytotechnol 15:145-155, 1994). With either protocol, an expressed, secreted heterologous protein can be repeatedly isolated from the cell culture supernatant, lysate, or membrane fractions depending on the disposition of the expressed protein in the cell. Within the infected 293 cell production protocol, non-secreted proteins may also be effectively obtained.

Assays can be used to measure other cellular responses, that include, chemotaxis, adhesion, changes in ion channel influx, regulation of second messenger levels and neurotransmitter release. Such assays are well known in the art. See, for example, in "Basic & Clinical Endocrinology Ser., Vol. Vol. 3," Cytochemical Bioassays: Techniques & Applications, Chayen; Chayen, Bitensky, eds., Dekker, New York, 1983.

In view of the tissue distribution observed for this zCTGF4, agonists (including the natural ligand) and antagonists have enormous potential in both *in vitro* and *in vivo* applications. Compounds identified as zCTGF4

agonists are useful for stimulating proliferation and/or differentiation of cells in culture. For example, agonist compounds are useful as components of defined cell culture media, and may be used alone or in combination with other 5 cytokines and hormones to replace serum that is commonly used in cell culture. Agonists are thus useful in specifically promoting the growth and/or development of cells derived from testis, trachea, bone marrow or kidney tissues or endothelial and ligament-derived fibroblast 10 cells in culture.

Agonists (including zCTGF4) will be useful for increasing production of extracellular matrix components, and may be used in the treatment of connective tissue. Particularly, agonists will be useful as treatment for 15 ligaments, cartilage and tendons. By virtue of the tissue distribution for the expression of these molecules, agonists will be useful for enhancing healing and stabilizing wounds and as a component artificial skin. The presence of zCTGF4 expression in the bone marrow 20 suggests that the molecules of the present invention play a role in hematopoiesis. That role is likely indirect, where stromal cells within the architecture of the bone marrow secrete zCTGF4, modulating production of cells of the hematopoietic lineage.

25 Antagonists will be useful for inhibiting expression of specialized cell functions, such as production of extracellular components and inhibition of cell proliferation. Genes encoding polypeptides having potential zCTGF4 polypeptide binding domains can be 30 obtained by screening random peptide libraries displayed on phage (phage display) or on bacteria, such as *E. coli*. Nucleotide sequences encoding the polypeptides can be obtained in a number of ways, such as through random 35 mutagenesis and random polynucleotide synthesis. These random peptide display libraries can be used to screen for peptides which interact with a known target which can be a

protein or polypeptide, such as a ligand or receptor, a biological or synthetic macromolecule, or organic or inorganic substances. Techniques for creating and screening such random peptide display libraries are known 5 in the art (Ladner et al., U.S. Patent 5,223,409; Ladner et al., U.S. Patent 4,946,778; Ladner et al., U.S. Patent 5,403,484 and Ladner et al., U.S. Patent 5,571,698) and random peptide display libraries and kits for screening such libraries are available commercially, for instance 10 from Clontech (Palo Alto, CA), Invitrogen Inc. (San Diego, CA), New England Biolabs, Inc. (Beverly, MA) and Pharmacia LKB Biotechnology Inc. (Piscataway, NJ). Random peptide display libraries can be screened using the zCTGF4 sequences disclosed herein to identify proteins which bind 15 to zCTGF4. These "binding proteins" which interact with zCTGF4 polypeptides may be used for tagging cells; for isolating homolog polypeptides by affinity purification; they can be directly or indirectly conjugated to drugs, toxins, radionuclides and the like. These binding 20 proteins can also be used in analytical methods such as for screening expression libraries and neutralizing activity. The binding proteins can also be used for diagnostic assays for determining circulating levels of polypeptides; for detecting or quantitating soluble 25 polypeptides as marker of underlying pathology or disease. These binding proteins can also act as zCTGF4 "antagonists" to block zCTGF4 binding and signal transduction *in vitro* and *in vivo*. These anti- zCTGF4 binding proteins would be useful for inhibiting expression 30 of genes which result in proliferation, differentiation or induction of specialized cell functions, such as production of extracellular matrix. Such anti-zCTGF4 binding proteins can be used for treatment in bone marrow fibrosis, modulating production or differentiation of 35 hematopoietic cells, prevention of scar tissue formation, cutaneous lupus erythematosis, scleroderma, dermatositis,

and end-stage kidney failure, alone or combination with other therapies.

zCTGF4 can also be used to identify inhibitors (antagonists) of its activity. Test compounds are added 5 to the assays disclosed herein to identify compounds that inhibit the activity of zCTGF4. In addition to those assays disclosed herein, samples can be tested for inhibition of zCTGF4 activity within a variety of assays designed to measure receptor binding or the 10 stimulation/inhibition of zCTGF4-dependent cellular responses. For example, zCTGF4-responsive cell lines can be transfected with a reporter gene construct that is responsive to a zCTGF4-stimulated cellular pathway. Reporter gene constructs of this type are known in the 15 art, and will generally comprise a zCTGF4-DNA response element operably linked to a gene encoding an assayable protein, such as luciferase. DNA response elements can include, but are not limited to, cyclic AMP response elements (CRE), hormone response elements (HRE) insulin 20 response element (IRE) (Nasrin et al., Proc. Natl. Acad. Sci. USA 87:5273-7, 1990) and serum response elements (SRE) (Shaw et al. Cell 56: 563-72, 1989). Cyclic AMP response elements are reviewed in Roestler et al., J. Biol. Chem. 263 (19):9063-6; 1988 and Habener, Molec. Endocrinol. 4 (8):1087-94; 1990. Hormone response 25 elements are reviewed in Beato, Cell 56:335-44; 1989. Candidate compounds, solutions, mixtures or extracts are tested for the ability to inhibit the activity of zCTGF4 on the target cells as evidenced by a decrease in zCTGF4 30 stimulation of reporter gene expression. Assays of this type will detect compounds that directly block zCTGF4 binding to cell-surface receptors, as well as compounds that block processes in the cellular pathway subsequent to receptor-ligand binding. In the alternative, compounds or 35 other samples can be tested for direct blocking of zCTGF4 binding to receptor using zCTGF4 tagged with a detectable label (e.g., ¹²⁵I, biotin, horseradish peroxidase, FITC,

and the like). Within assays of this type, the ability of a test sample to inhibit the binding of labeled zCTGF4 to the receptor is indicative of inhibitory activity, which can be confirmed through secondary assays. Receptors used 5 within binding assays may be cellular receptors or isolated, immobilized receptors.

A ZCTGF4 polypeptide can be expressed as a fusion with an immunoglobulin heavy chain constant region, typically an F_C fragment, which contains two constant 10 region domains and lacks the variable region. Methods for preparing such fusions are disclosed in U.S. Patents Nos. 5,155,027 and 5,567,584. Such fusions are typically secreted as multimeric molecules wherein the Fc portions are disulfide bonded to each other and two non-Ig 15 polypeptides are arrayed in closed proximity to each other. Fusions of this type can be used to affinity purify ligand, as an *in vitro* assay tool, or antagonist. For use in assays, the chimeras are bound to a support via the F_C region and used in an ELISA format.

20 A zCTGF4 ligand-binding polypeptide can also be used for purification of ligand. The polypeptide is immobilized on a solid support, such as beads of agarose, cross-linked agarose, glass, cellulosic resins, silica-based resins, polystyrene, cross-linked polyacrylamide, or 25 like materials that are stable under the conditions of use. Methods for linking polypeptides to solid supports are known in the art, and include amine chemistry, cyanogen bromide activation, N-hydroxysuccinimide activation, epoxide activation, sulfhydryl activation, and 30 hydrazide activation. The resulting medium will generally be configured in the form of a column, and fluids containing ligand are passed through the column one or more times to allow ligand to bind to the receptor polypeptide. The ligand is then eluted using changes in 35 salt concentration, chaotropic agents (guanidine HCl), or pH to disrupt ligand-receptor binding.

An assay system that uses a ligand-binding receptor (or an antibody, one member of a complement/anti-complement pair) or a binding fragment thereof, and a commercially available biosensor instrument (BIAcore, 5 Pharmacia Biosensor, Piscataway, NJ) may be advantageously employed. Such receptor, antibody, member of a complement/anti-complement pair or fragment is immobilized onto the surface of a receptor chip. Use of this instrument is disclosed by Karlsson, J. Immunol. Methods 10 145:229-40, 1991 and Cunningham and Wells, J. Mol. Biol. 234:554-63, 1993. A receptor, antibody, member or fragment is covalently attached, using amine or sulphhydryl chemistry, to dextran fibers that are attached to gold film within the flow cell. A test sample is passed 15 through the cell. If a ligand, epitope, or opposite member of the complement/anti-complement pair is present in the sample, it will bind to the immobilized receptor, antibody or member, respectively, causing a change in the refractive index of the medium, which is detected as a 20 change in surface plasmon resonance of the gold film. This system allows the determination of on- and off-rates, from which binding affinity can be calculated, and assessment of stoichiometry of binding.

Ligand-binding receptor polypeptides can also be 25 used within other assay systems known in the art. Such systems include Scatchard analysis for determination of binding affinity (see Scatchard, Ann. NY Acad. Sci. 51: 660-72, 1949) and calorimetric assays (Cunningham et al., Science 253:545-48, 1991; Cunningham et al., Science 30 245:821-25, 1991).

ZCTGF4 polypeptides can also be used to prepare antibodies that bind to zCTGF4 epitopes, peptides or polypeptides. The zCTGF4 polypeptide or a fragment thereof serves as an antigen (immunogen) to inoculate an animal 35 and elicit an immune response. One of skill in the art would recognize that antigenic, epitope-bearing

polypeptides contain a sequence of at least 6, preferably at least 9, and more preferably at least 15 to about 30 contiguous amino acid residues of a zCTGF4 polypeptide (e.g., SEQ ID NO:2). Polypeptides comprising a larger portion of a zCTGF4 polypeptide, i.e., from 30 to 10 residues up to the entire length of the amino acid sequence are included. Antigens or immunogenic epitopes can also include attached tags, adjuvants and carriers, as described herein. Suitable antigens include the zCTGF4 polypeptide encoded by SEQ ID NO:2 from amino acid number 24 to amino acid number 354, or a contiguous 9 to 330 amino acid fragment thereof. Other suitable antigens include the IBP domain (residues 59-102), von Willebrand factor c domain (residues 114-179), variable domain (residues 180-208), and sulfated glyconconjugate binding motif domain (residues 209-252) as disclosed herein. Preferred peptides to use as antigens are hydrophilic peptides such as those predicted by one of skill in the art from a hydrophobicity plot (See Figure). ZCTGF4 hydrophilic peptides include peptides comprising amino acid sequences selected from the group consisting of: residues 239-244, residues 105-110, residues 172-177, residues 238-243 and residues 171-176, all of SEQ ID NO:2. Antibodies from an immune response generated by inoculation of an animal with these antigens can be isolated and purified as described herein. Methods for preparing and isolating polyclonal and monoclonal antibodies are well known in the art. See, for example, Current Protocols in Immunology, Cooligan, et al. (eds.), National Institutes of Health, John Wiley and Sons, Inc., 1995; Sambrook et al., Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor, NY, 1989; and Hurrell, J. G. R., Ed., Monoclonal Hybridoma Antibodies:

Techniques and Applications, CRC Press, Inc., Boca Raton, FL, 1982.

As would be evident to one of ordinary skill in the art, polyclonal antibodies can be generated from 5 inoculating a variety of warm-blooded animals such as horses, cows, goats, sheep, dogs, chickens, rabbits, mice, and rats with a zCTGF4 polypeptide or a fragment thereof.

The immunogenicity of a zCTGF4 polypeptide may be increased through the use of an adjuvant, such as alum 10 (aluminum hydroxide) or Freund's complete or incomplete adjuvant. Polypeptides useful for immunization also include fusion polypeptides, such as fusions of zCTGF4 or a portion thereof with an immunoglobulin polypeptide or with maltose binding protein. The polypeptide immunogen 15 may be a full-length molecule or a portion thereof. If the polypeptide portion is "hapten-like", such portion may be advantageously joined or linked to a macromolecular carrier (such as keyhole limpet hemocyanin (KLH), bovine serum albumin (BSA) or tetanus toxoid) for immunization.

20 As used herein, the term "antibodies" includes polyclonal antibodies, affinity-purified polyclonal antibodies, monoclonal antibodies, and antigen-binding fragments, such as $F(ab')_2$ and Fab proteolytic fragments. Genetically engineered intact antibodies or fragments, 25 such as chimeric antibodies, Fv fragments, single chain antibodies and the like, as well as synthetic antigen-binding peptides and polypeptides, are also included. Non-human antibodies may be humanized by grafting non-human CDRs onto human framework and constant regions, or 30 by incorporating the entire non-human variable domains (optionally "cloaking" them with a human-like surface by replacement of exposed residues, wherein the result is a "veeneered" antibody). In some instances, humanized antibodies may retain non-human residues within the human

variable region framework domains to enhance proper binding characteristics. Through humanizing antibodies, biological half-life may be increased, and the potential for adverse immune reactions upon administration to humans
5 is reduced. Moreover, human antibodies can be produced in transgenic, non-human animals that have been engineered to contain human immunoglobulin genes as disclosed in WIPO Publication WO 98/24893. It is preferred that the endogenous immunoglobulin genes in these animals be
10 inactivated or eliminated, such as by homologous recombination.

Antibodies are considered to be specifically binding if: 1) they exhibit a threshold level of binding activity, and 2) they do not significantly cross-react
15 with related polypeptide molecules. A threshold level of binding is determined if anti- zCTGF4 antibodies herein bind to a zCTGF4 polypeptide, peptide or epitope with an affinity at least 10-fold greater than the binding affinity to control (non- zCTGF4) polypeptide. It is
20 preferred that the antibodies exhibit a binding affinity (K_a) of 10^6 M^{-1} or greater, preferably 10^7 M^{-1} or greater,
 10^8 M^{-1} or greater, and most preferably
 10^9 M^{-1} or greater. The binding affinity of an antibody
25 can be readily determined by one of ordinary skill in the art, for example, by Scatchard analysis (Scatchard, G.,
Ann. NY Acad. Sci. 51: 660-672, 1949).

Whether anti-zCTGF4 antibodies do not significantly cross-react with related polypeptide molecules is shown, for example, by the antibody detecting
30 zCTGF4 polypeptide but not known related polypeptides using a standard Western blot analysis (Ausubel et al.,
ibid.). Examples of known related polypeptides are those disclosed in the prior art, such as known orthologs, and

paralogs, and similar known members of a protein family, such as other known human CTGFs (e.g., CTGF and CTGF-2). Screening can also be done using non-human zCTGF4, and zCTGF4 mutant polypeptides. Moreover, antibodies can be 5 "screened against" known related polypeptides, to isolate a population that specifically binds to the zCTGF4 polypeptides. For example, antibodies raised to zCTGF4 are adsorbed to related polypeptides adhered to insoluble matrix; antibodies specific to zCTGF4 will flow through 10 the matrix under the proper buffer conditions. Screening allows isolation of polyclonal and monoclonal antibodies non-crossreactive to known closely related polypeptides (Antibodies: A Laboratory Manual, Harlow and Lane (eds.), Cold Spring Harbor Laboratory Press, 1988; Current 15 Protocols in Immunology, Cooligan, et al. (eds.), National Institutes of Health, John Wiley and Sons, Inc., 1995). Screening and isolation of specific antibodies is well known in the art. See, Fundamental Immunology, Paul (eds.), Raven Press, 1993; Getzoff et al., Adv. in 20 Immunol. 43: 1-98, 1988; Monoclonal Antibodies: Principles and Practice, Goding, J.W. (eds.), Academic Press Ltd., 1996; Benjamin et al., Ann. Rev. Immunol. 2: 67-101, 1984. Specifically binding anti-zCTGF4 antibodies can be detected by a number of methods in the art, and 25 disclosed below.

A variety of assays known to those skilled in the art can be utilized to detect antibodies which bind to zCTGF4 proteins or polypeptides. Exemplary assays are described in detail in Antibodies: A Laboratory Manual, 30 Harlow and Lane (Eds.), Cold Spring Harbor Laboratory Press, 1988. Representative examples of such assays include: concurrent immunoelectrophoresis, radioimmunoassay, radioimmuno-precipitation, enzyme-linked immunosorbent assay (ELISA), dot blot or Western blot

assay, inhibition or competition assay, and sandwich assay. In addition, antibodies can be screened for binding to wild-type versus mutant zCTGF4 polypeptide.

Alternative techniques for generating or selecting antibodies useful herein include *in vitro* exposure of lymphocytes to zCTGF4 protein or peptide, and selection of antibody display libraries in phage or similar vectors (for instance, through use of immobilized or labeled zCTGF4 protein or peptide). Genes encoding polypeptides having potential zCTGF4 polypeptide binding domains can be obtained by screening random peptide libraries displayed on phage (phage display) or on bacteria, such as *E. coli*. Nucleotide sequences encoding the polypeptides can be obtained in a number of ways, such as through random mutagenesis and random polynucleotide synthesis. These random peptide display libraries can be used to screen for peptides which interact with a known target which can be a protein or polypeptide, such as a ligand or receptor, a biological or synthetic macromolecule, or organic or inorganic substances. Techniques for creating and screening such random peptide display libraries are known in the art (Ladner et al., US Patent NO. 5,223,409; Ladner et al., US Patent NO. 4,946,778; Ladner et al., US Patent NO. 5,403,484 and Ladner et al., US Patent NO. 5,571,698) and random peptide display libraries and kits for screening such libraries are available commercially, for instance from Clontech (Palo Alto, CA), Invitrogen Inc. (San Diego, CA), New England Biolabs, Inc. (Beverly, MA) and Pharmacia LKB Biotechnology Inc. (Piscataway, NJ). Random peptide display libraries can be screened using the zCTGF4 sequences disclosed herein to identify proteins which bind to zCTGF4. These "binding polypeptides" which interact with zCTGF4 polypeptides can be used for tagging cells;

for isolating homolog polypeptides by affinity purification; they can be directly or indirectly conjugated to drugs, toxins, radionuclides and the like. These binding polypeptides can also be used in analytical methods such as for screening expression libraries and neutralizing activity, e.g., for blocking interaction between ligand and receptor, or viral binding to a receptor. The binding polypeptides can also be used for diagnostic assays for determining circulating levels of zCTGF4 polypeptides; for detecting or quantitating soluble zCTGF4 polypeptides as marker of underlying pathology or disease. These binding polypeptides can also act as zCTGF4 "antagonists" to block zCTGF4 binding and signal transduction *in vitro* and *in vivo*. These anti-zCTGF4 binding polypeptides would be useful for inhibiting zCTGF4 activity or protein-binding.

Antibodies to zCTGF4 may be used for tagging cells that express zCTGF4; for isolating zCTGF4 by affinity purification; for diagnostic assays for determining circulating levels of zCTGF4 polypeptides; for detecting or quantitating soluble zCTGF4 as marker of underlying pathology or disease; in analytical methods employing FACS; for screening expression libraries; for generating anti-idiotypic antibodies; and as neutralizing antibodies or as antagonists to block zCTGF4 *in vitro* and *in vivo*. In particular, antibodies will be useful for diagnostics, due to the association of proteins of the present invention with extracellular matrix and vessels, and labeled proteins will be useful in the diagnosis of diseases such as bone marrow fibrosis, prevention of scar tissue formation, cutaneous lupus erythematosis, scleroderma, dermatositis, and end-stage kidney failure.

Antibodies or polypeptides herein may also be directly or indirectly conjugated to drugs, toxins,

radionuclides and the like, and these conjugates used for *in vivo* diagnostic or therapeutic applications. Moreover, antibodies to zCTGF4 or fragments thereof may be used *in vitro* to detect denatured zCTGF4 or fragments thereof in 5 assays, for example, Western Blots or other assays known in the art.

Antibodies or polypeptides herein can also be directly or indirectly conjugated to drugs, toxins, radionuclides and the like, and these conjugates used for 10 *in vivo* diagnostic or therapeutic applications. For instance, polypeptides or antibodies of the present invention can be used to identify or treat tissues or organs that express a corresponding anti-complementary molecule (receptor or antigen, respectively, for 15 instance). More specifically, zCTGF4 polypeptides or anti-zCTGF4 antibodies, or bioactive fragments or portions thereof, can be coupled to detectable or cytotoxic molecules and delivered to a mammal having cells, tissues or organs that express the anti-complementary molecule. 20 For example, for use of the antibodies and polypeptides, labeled for detection by imaging technologies, will be useful for diagnosing diseases associated with extracellular matrix and vessels, such as, bone marrow fibrosis, aberrant hematopoiesis, prevention of scar 25 tissue formation, cutaneous lupus erythematosis, scleroderma, dermatositis, and end-stage kidney failure.

Suitable detectable molecules may be directly or indirectly attached to the polypeptide or antibody, and include radionuclides, enzymes, substrates, cofactors, 30 inhibitors, fluorescent markers, chemiluminescent markers, magnetic particles and the like. Suitable cytotoxic molecules may be directly or indirectly attached to the polypeptide or antibody, and include bacterial or plant toxins (for instance, diphtheria toxin, *Pseudomonas* 35 exotoxin, ricin, abrin and the like), as well as therapeutic radionuclides, such as iodine-131, rhenium-188

or yttrium-90 (either directly attached to the polypeptide or antibody, or indirectly attached through means of a chelating moiety, for instance). Polypeptides or antibodies may also be conjugated to cytotoxic drugs, such 5 as adriamycin. For indirect attachment of a detectable or cytotoxic molecule, the detectable or cytotoxic molecule can be conjugated with a member of a complementary/ anticomplementary pair, where the other member is bound to the polypeptide or antibody portion. For these purposes, 10 biotin/streptavidin is an exemplary complementary/ anticomplementary pair.

In another embodiment, polypeptide-toxin fusion proteins or antibody-toxin fusion proteins can be used for targeted cell or tissue inhibition or ablation (for 15 instance, to treat diseases caused by inappropriate growth of cells or tissues). Such molecule fusion proteins thus represent a generic targeting vehicle for cell/tissue-specific delivery of generic anti-complementary-detectable/ cytotoxic molecule conjugates.

20 In another embodiment, zCTGF4-cytokine fusion proteins or antibody-cytokine fusion proteins can be used for enhancing *in vivo* killing of target tissues (for example, blood and bone marrow cancers), if the zCTGF4 polypeptide or anti-zCTGF4 antibody targets the 25 hyperproliferative blood or bone marrow cell (See, generally, Hornick et al., Blood 89:4437-47, 1997). They described fusion proteins enable targeting of a cytokine to a desired site of action, thereby providing an elevated local concentration of cytokine. Suitable zCTGF4 30 polypeptides or anti-zCTGF4 antibodies target an undesirable cell or tissue (i.e., a tumor or a leukemia), and the fused cytokine mediated improved target cell lysis by effector cells. Suitable cytokines for this purpose include interleukin 2 and granulocyte-macrophage colony- 35 stimulating factor (GM-CSF), for instance.

In another embodiment, if the zCTGF4 polypeptide or anti-zCTGF4 antibody targets vascular cells or tissues, such polypeptide or antibody may be conjugated with a radionuclide, and particularly with a beta-emitting radionuclide, to reduce restenosis. Such therapeutic approach poses less danger to clinicians who administer the radioactive therapy. For instance, iridium-192 impregnated ribbons placed into stented vessels of patients until the required radiation dose was delivered showed decreased tissue growth in the vessel and greater luminal diameter than the control group, which received placebo ribbons. Further, revascularisation and stent thrombosis were significantly lower in the treatment group. Similar results are predicted with targeting of a bioactive conjugate containing a radionuclide, as described herein.

The bioactive polypeptide or antibody conjugates described herein can be delivered intravenously, intraarterially or intraductally, or may be introduced locally at the intended site of action.

Molecules of the present invention can be used to identify and isolate receptors involved in growth and differentiation of zCTGF4 responsive cells. For example, proteins and peptides of the present invention can be immobilized on a column and membrane preparations run over the column (Immobilized Affinity Ligand Techniques, Hermanson et al., eds., Academic Press, San Diego, CA, 1992, pp.195-202). Proteins and peptides can also be radiolabeled (Methods in Enzymol., vol. 182, "Guide to Protein Purification", M. Deutscher, ed., Acad. Press, San Diego, 1990, 721-737) or photoaffinity labeled (Brunner et al., Ann. Rev. Biochem. 62:483-514, 1993 and Fedan et al., Biochem. Pharmacol. 33:1167-1180, 1984) and specific cell-surface proteins can be identified.

The molecules of the present invention will be useful for regulating the growth and/or differentiation of

zCTGF4 responsive cells. The polypeptides, nucleic acid and/or antibodies of the present invention can be used in treatment of disorders associated with unregulated growth in zCTGF4-responsive tissues. In particular, the 5 molecules of the present may be used to produce antagonists to treat or prevent development of pathological conditions in tissues as testis, trachea, bone marrow, and kidney. Certain diseases such as bone marrow fibrosis, prevention of scar tissue formation, cutaneous lupus erythematosis, 10 scleroderma, dermatositis, and end-stage kidney failure, may be amenable to such diagnosis, treatment or prevention.

Polynucleotides encoding zCTGF4 polypeptides are useful within gene therapy applications where it is 15 desired to increase or inhibit zCTGF4 activity. If a mammal has a mutated or absent zCTGF4 gene, the zCTGF4 gene can be introduced into the cells of the mammal. In one embodiment, a gene encoding a zCTGF4 polypeptide is introduced *in vivo* in a viral vector. Such vectors 20 include an attenuated or defective DNA virus, such as, but not limited to, herpes simplex virus (HSV), papillomavirus, Epstein Barr virus (EBV), adenovirus, adeno-associated virus (AAV), and the like. Defective viruses, which entirely or almost entirely lack viral 25 genes, are preferred. A defective virus is not infective after introduction into a cell. Use of defective viral vectors allows for administration to cells in a specific, localized area, without concern that the vector can infect other cells. Examples of particular vectors include, but 30 are not limited to, a defective herpes simplex virus 1 (HSV1) vector (Kaplitt et al., Molec. Cell. Neurosci. 2:320-30, 1991); an attenuated adenovirus vector, such as the vector described by Stratford-Perricaudet et al., J. Clin. Invest. 90:626-30, 1992; and a defective adeno- 35 associated virus vector (Samulski et al., J. Virol.

61:3096-101, 1987; Samulski et al., J. Virol. 63:3822-28, 1989).

In another embodiment, the zCTGF4 gene can be introduced in a retroviral vector, e.g., as described in 5 Anderson et al., U.S. Patent No. 5,399,346; Mann et al. Cell 33:153, 1983; Temin et al., U.S. Patent No. 4,650,764; Temin et al., U.S. Patent No. 4,980,289; 10 Markowitz et al., J. Virol. 62:1120, 1988; Temin et al., U.S. Patent No. 5,124,263; International Patent Publication No. WO 95/07358, published March 16, 1995 by Dougherty et al.; and Kuo et al., Blood 82:845-852, 1993. Alternatively, the vector can be introduced by lipofection 15 in vivo using liposomes. Synthetic cationic lipids can be used to prepare liposomes for in vivo transfection of a gene encoding a marker (Felgner et al., Proc. Natl. Acad. Sci. USA 84:7413-17, 1987; Mackey et al., Proc. Natl. Acad. Sci. USA 85:8027-31, 1988). The use of lipofection 20 to introduce exogenous genes into specific organs in vivo has certain practical advantages. Molecular targeting of 25 liposomes to specific cells represents one area of benefit. More particularly, directing transfection to particular cells represents one area of benefit. For instance, directing transfection to particular cell types would be particularly advantageous in a tissue with cellular heterogeneity, such as the pancreas, liver, kidney, and brain. Lipids may be chemically coupled to 30 other molecules for the purpose of targeting. Targeted peptides (e.g., hormones or neurotransmitters), proteins such as antibodies, or non-peptide molecules can be coupled to liposomes chemically.

It is possible to remove the target cells from the body; to introduce the vector as a naked DNA plasmid; and then to re-implant the transformed cells into the body. Naked DNA vectors for gene therapy can be 35 introduced into the desired host cells by methods known in the art, e.g., transfection, electroporation,

microinjection, transduction, cell fusion, DEAE dextran, calcium phosphate precipitation, use of a gene gun or use of a DNA vector transporter. See, e.g., Wu et al., J. Biol. Chem. 267:963-67, 1992; Wu et al., J. Biol. Chem. 5 263:14621-24, 1988.

Antisense methodology can be used to inhibit zCTGF4 gene transcription, such as to inhibit cell proliferation in vivo. Polynucleotides that are complementary to a segment of a zCTGF4-encoding 10 polynucleotide (e.g., a polynucleotide as set forth in SEQ ID NO:1) are designed to bind to zCTGF4-encoding mRNA and to inhibit translation of such mRNA. Such antisense polynucleotides are used to inhibit expression of zCTGF4 polypeptide-encoding genes in cell culture or in a 15 subject.

The present invention also provides reagents which will find use in diagnostic applications. For example, the zCTGF4 gene, a probe comprising zCTGF4 DNA or RNA or a subsequence thereof can be used to determine if 20 the zCTGF4 gene is present on chromosome 6 or if a mutation has occurred. Detectable chromosomal aberrations at the zCTGF4 gene locus include, but are not limited to, aneuploidy, gene copy number changes, insertions, deletions, restriction site changes and rearrangements. 25 Such aberrations can be detected using polynucleotides of the present invention by employing molecular genetic techniques, such as restriction fragment length polymorphism (RFLP) analysis, short tandem repeat (STR) analysis employing PCR techniques, and other genetic 30 linkage analysis techniques known in the art (Sambrook et al., ibid.; Ausubel et. al., ibid.; Marian, Chest 108:255-65, 1995).

Mice engineered to express the zCTGF4 gene, referred to as "transgenic mice," and mice that exhibit a 35 complete absence of zCTGF4 gene function, referred to as "knockout mice," may also be generated (Snouwaert et al.,

Science 257:1083, 1992; Lowell et al., Nature 366:740-42, 1993; Capecchi, M.R., Science 244: 1288-1292, 1989; Palmiter, R.D. et al. Annu Rev Genet. 20: 465-499, 1986). For example, transgenic mice that over-express zCTGF4, either ubiquitously or under a tissue-specific or tissue-restricted promoter can be used to ask whether over-expression causes a phenotype. For example, over-expression of a wild-type zCTGF4 polypeptide, polypeptide fragment or a mutant thereof may alter normal cellular processes, resulting in a phenotype that identifies a tissue in which zCTGF4 expression is functionally relevant and may indicate a therapeutic target for the zCTGF4, its agonists or antagonists. Moreover, such over-expression may result in a phenotype that shows similarity with human diseases. Similarly, knockout zCTGF4 mice can be used to determine where zCTGF4 is absolutely required *in vivo*. The phenotype of knockout mice is predictive of the *in vivo* effects of that a zCTGF4 antagonist, such as those described herein, may have. The human zCTGF4 cDNA can be used to isolate murine zCTGF4 mRNA, cDNA and genomic DNA, which are subsequently used to generate knockout mice. These mice may be employed to study the zCTGF4 gene and the protein encoded thereby in an *in vivo* system, and can be used as *in vivo* models for corresponding human diseases. Moreover, transgenic mice expression of zCTGF4 antisense polynucleotides or ribozymes directed zCTGF4, described herein, can be used analogously to transgenic mice described above.

For pharmaceutical use, the proteins of the present invention are formulated for parenteral, particularly intravenous or subcutaneous, delivery according to conventional methods. Intravenous administration will be by bolus injection or infusion over a typical period of one to several hours. In general,

pharmaceutical formulations will include a zCTGF4 protein in combination with a pharmaceutically acceptable vehicle, such as saline, buffered saline, 5% dextrose in water or the like. Formulations may further include one or more 5 excipients, preservatives, solubilizers, buffering agents, albumin to prevent protein loss on vial surfaces, etc. Methods of formulation are well known in the art and are disclosed, for example, in Remington: The Science and Practice of Pharmacy, Gennaro, ed., Mack Publishing Co., 10 Easton, PA, 19th ed., 1995. Therapeutic doses will generally be in the range of 0.1 to 100 µg/kg of patient weight per day, preferably 0.5-20 µg/kg per day, with the exact dose determined by the clinician according to accepted standards, taking into account the nature and 15 severity of the condition to be treated, patient traits, etc. Determination of dose is within the level of ordinary skill in the art. The proteins may be administered for acute treatment, over one week or less, often over a period of one to three days or may be used in 20 chronic treatment, over several months or years. The invention is further illustrated by the following non-limiting examples.

The invention is further illustrated by the following non-limiting examples.

25

EXAMPLES

Example 1

30 Scanning of a translated DNA database using the CTGF family motifs as a query resulted in identification of an EST sequence found to have some homology to the connective tissue growth factor family.

Plasmid DNA was isolated from a clone 35 corresponding to the EST, which had been designated zCTGF4, and was analyzed for polynucleotide sequence.

Upon alignment of the zCTGF4 cDNA and CTGF family, it was revealed that the clone contained a truncated cDNA sequence with an intron at the 5' end of the sequence.

Based on the tissue distribution from Northern blot analysis (see Example 2), a cDNA library was constructed from human testis, and used to screen for a full length clone. The library was calculated to contain approximately 10^6 clones. The master plate containing 80 pools (each pool representing 12 pools of 1.25×10^4 clones) was screened using PCR to determine whether zCTGF4 cDNA was present. Reactions were set up using: 1 μ l of each pool, 20 pmoles each of oligonucleotide primers ZC14,882 and ZC14,883 (SEQ ID NOS: 6 and 7, respectively), and 1 U of *ExTaq*® DNA polymerase (TaKaRa Shuzo Co., Ltd., Shiga, JP) in a 25 μ l volume. PCR was performed in a 96-well plate on a GenAmp PCR system 9700 (PE Applied Biosystems, Cheshire, UK). The reaction was run as follows: 94°C for 1.5 minute, then for 30 cycles of 94°C, 15 seconds; 55°C 20 seconds; 72°C 30 seconds; and ended with a 7 minute incubation at 72°C. Pools designated A3, A6, D3, D6 and F7 were positive for the presence of zCTGF4 DNA, and selected for further analyses.

Positive pools were further analyzed with PCR to identify clones with longer 5' end sequence. The reaction mixture contained: 1 μ l of each pool, 20 pmoles each of oligonucleotide primers ZC15,909 (corresponding to a vector sequence) and ZC14,885 (corresponding to a gene specific sequence) (SEQ ID NOS: 8 and 9, respectively), and 1 U of mixture *ExTaq* (TaKaRa) and *Pfu*® (Stratagene, La Jolla, CA) (2:1) DNA polymerase, in a 25 μ l total volume. PCR was performed on a GenAmp PCR system 2400 (PE Biosystems) as follows: 94°C for 1.5 minute, then for 25 cycles of 94°C, 15 seconds; 55°C 20 seconds; 72°C 30 seconds; 7 minute incubation at 72°C. A second nested anchor PCR was performed using 1 μ l of 1/50 diluted first

round PCR products as template, 20 pmoles each of oligonucleotide primers ZC15,911 (corresponding to vector sequence) and ZC14,884 (corresponding to gene specific sequence) (SEQ ID NOS: 10 and 11, respectively), and 1 U 5 of mixture *ExTaq* (TaKaRa) and *Pfu*® (Stratagene) (2:1) DNA polymerase in a 25 µl total volume. The reaction was run as follows: 94°C for 1.5 minute, then for 25 cycles of 94°C, 15 seconds; 50°C 20 seconds; 72°C 30 seconds; 7 minute incubation at 72°C. PCR products from A3, A6, D3 10 and D6 were gel purified with QIAquick Gel Extraction kit (Qiagen Inc. Chatsworth, CA), subcloned into a pCR2.1 vector of TA Cloning® kit (Invitrogen, Carlsbad, CA), which provides for direct ligation into expression vectors 15 (Mead et al., Bio/Technology 9(7):657-663, 1991), and designated as CTGF4a3, CTGF4a6, CTGF4d3, CTGF4d6.

Sequence analysis of CTGF4a3, CTGF4a6, CTGF4d3, and CTGF4d6 revealed that the sequence of CTGF4a3 encoded a start Met with a putative signal peptide, a long open reading frame (ORF) at which the first reading frame was 20 interrupted by a stop codon, and a short ORF thereafter at the second reading frame. The short ORF was identical to the 5' end coding region of zCTGF4, while the long ORF had a 45% sequence similarity to human CTGF. The sequence of CTGF4a6 was almost identical to CTGF4a3 except it was 25 short 16 N-terminal amino acids in the secretory signal peptide. Both CTGF4d3 and CTGF4d6 had smaller inserts. All four clones encoded a long ORF interrupted by a stop codon at same place.

A PCR approach, as described previously, was 30 used to find a correctly coded sequence in other tissues. A human bone marrow marathon cDNA was used based on the results of northern blot analysis (see Example 2). cDNA was made using Clontech Marathon™ cDNA kit (Clontech, Palo Alto, CA), according to the manufacturer's 35 specifications. 5 µl of 1/100 diluted Marathon™ bone marrow cDNA, 20 pmoles each of oligonucleotide primers

ZC15,910 and ZC14,885 (SEQ ID NOS: 12 and 9, respectively), and 1 U of *ExTaq* (TaKaRa) and *Pfu*® (Stratagene) (2:1) DNA polymerase were used in a 25 µl of reactions. The reactions were run as follows: 94°C for 5 1.5 minute, then for 25 cycles of 94°C, 15 seconds; 54°C 20 seconds; 72°C 30 seconds; and ended with a 7 minute incubation at 72°C. 1 µl of 1/50 diluted first PCR product was used as template for nested PCR. 20 pmoles each of oligonucleotide primers ZC16,192 and ZC14,884 (SEQ 10 ID NOS: 13 and 11, respectively), and 1 U of *ExTaq* (TaKaRa) and *Pfu*® (Stratagene) (2:1) DNA polymerase were used in a 25 µl of reactions. The reactions were run as follows: 94°C for 1.5 minute, then for 25 cycles of 94°C, 15 seconds; 50°C 20 seconds; 72°C 30 seconds; and ended 15 with a 7 minute incubation at 72°C. The PCR product was gel purified, subcloned into pCR2.1 vector (as described previously), and sequenced. The sequence analysis revealed that the 5' and 3' end of bone marrow clone overlapped with the testis clones, with a discrepancy in 20 the region having the reading frame shift in the testis clone. The sequence of bone marrow clone had a correct ORF, and was included in the composite full length sequence of zCTGF4. The initially identified EST, which originated from a penis library, as well as clones 25 identified from two testis libraries, had introns or reading frame shifts in the same region. Further analysis revealed that this region was least conserved area when compared to other members in the family, suggesting the reading frame shift may be a regulatory mechanism.

30

Example 2

Northern analyses were performed using Human Multiple Tissue Blots I, II and III from Clontech (Palo Alto, CA). A probe was generated from a gel purified PCR 35 product made from ZC14,883 (SEQ ID NO: 7) and ZC14,882

(SEQ ID NO: 6) as primers and zCTGF4 as template, that had been radioactively labeled with REDIPRIME™ DNA labeling kit (Amersham, Arlington Heights, IL) according to the manufacturer's suggestion. The probe was purified using a 5 NUCTRAP push column (Stratagene). EXPRESSHYB™ (Clontech) solution was used for prehybridization and as a hybridizing solution for the Northern blots. Hybridization took place overnight at 65°C, and the blots were then washed in 2X SSC and 0.05% SDS at RT, followed 10 by a wash in 0.1X SSC and 0.1% SDS at 50°C. One major transcript was observed at size of 1.4 kb. Signals were present in testis, bone marrow, trachea, kidney, liver, stomach, small intestine, ovary, placenta, prostate and spinal cord. The expression of zCTGF4 was also examined 15 with Human RNA Master blot (Clontech) with a probe generated from a PCR product amplified with ZC16,192 (SEQ ID NO: 13) and ZC14,884 (SEQ ID NO: 11) as primers and zCTGF4 as template DNA. The conditions for probe labeling and hybridization were the same as described above. 20 zCTGF4 was positive in two other tissues, mammary gland and fetal kidney, in addition to those described above.

Example 3

25 zCTGF4 was mapped to chromosome 6 using the commercially available "GeneBridge 4 Radiation Hybrid Panel" (Research Genetics, Inc., Huntsville, AL). The GeneBridge 4 Radiation Hybrid Panel contained DNAs from each of 93 radiation hybrid clones, plus two control DNAs 30 (the HFL donor and the A23 recipient). A publicly available WWW server (<http://www-genome.wi.mit.edu/cgi-bin/contig/rhmapper.pl>) allowed mapping relative to the Whitehead Institute/MIT Center for Genome Research's radiation hybrid map of the human genome (the "WICGR" 35 radiation hybrid map) which was constructed with the GeneBridge 4 Radiation Hybrid Panel.

For the mapping of zCTGF4 with the "GeneBridge 4 RH Panel", 20 μ l reactions were set up in a 96-well microtiter plate (Stratagene, La Jolla, CA), and used in a "RoboCycler Gradient 96" thermal cycler (Stratagene).
5 Each of the 95 PCR reactions consisted of 2 μ l 10X KlenTaq PCR reaction buffer (CLONTECH Laboratories, Inc., Palo Alto, CA), 1.6 μ l dNTPs mix (2.5 mM each, PERKIN-ELMER, Foster City, CA), 1 μ l sense primer, ZC15,089 (SEQ ID NO: 14), 1 μ l antisense primer, ZC15,092 (SEQ ID NO: 15), 2 μ l
10 "RediLoad" (Research Genetics, Inc., Huntsville, AL), 0.4 μ l 50X Advantage KlenTaq Polymerase Mix (Clontech Laboratories, Inc.), 25 ng of DNA from an individual hybrid clone or control and ddH₂O for a total volume of 20 μ l. The reactions were overlaid with an equal amount of
15 mineral oil and sealed. The PCR cycler conditions were as follows: an initial 1 cycle 5 minute denaturation at 95°C, 35 cycles of a 1 minute denaturation at 95°C, 1 minute annealing at 56°C and 1.5 minute extension at 72°C, followed by a final 1 cycle extension of 7 minutes at
20 72°C. The reactions were separated by electrophoresis on a 2% agarose gel (GIBCO-BRL Life Technologies, Gaithersburg, MD).

The results showed that zCTGF4 maps 6.29 cR_3000 from the framework marker WI- 4792 on the WICGR chromosome
25 6 radiation hybrid map. Proximal and distal framework markers were WI-4792 and CHLC.GATA31.100, respectively. The use of surrounding markers positions zCTGF4 in the 6q22.1 region on the integrated LDB chromosome 6 map (The Genetic Location Database, University of Southampton, WWW server: http://cedar.genetics.soton.ac.uk/public_html/).

Example 4

Several expression constructs were made for eukaryotic expression of the zCTGF4 cDNA. A mammalian expression vector was constructed with the dihydrofolate reductase gene under control of the SV40 early promoter, SV40 polyadenylation site, a cloning site to insert the

gene of interest under control of the MT-1 promoter and the hGH polyadenylation site. The expression vector was designated pZP9 and is deposited at the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD.

5 To facilitate purification, pZP9 vector was modified by addition of the tPA leader sequence (U.S. Patent 5,641,655, incorporated herein by reference) and a GluGlu tag (SEQ ID NO: 16) between the MT-1 promoter and hGH terminator. The tPA leader replaces the native secretory

10 signal sequence for DNAs encoding for polypeptides of interest that are inserted into this vector, and expression results in an N-terminally tagged protein. The N-terminally tagged vector was designated pZP9NEE. Another vector was similarly constructed with a C-terminal

15 GluGlu tag (SEQ ID NO: 16) inserted just 5' to the hGH terminator and utilizes the native (or other fused) secretory signal sequence for secretion of the encoded polypeptide of interest, and expression resulted in a C-terminally tagged protein. The C-terminal GluGlu tagged

20 vector was designated pZP9CEE.

A 5' DNA fragment containing the zCTGF4 coding region (nucleotide 86 to nucleotide 613 of SEQ ID NO: 1) was generated by PCR using oligonucleotide primers ZC16,422 (SEQ ID NO: 17) and ZC16,424 (SEQ ID NO: 18) and CTGF4A3 as the template, and a 3' DNA fragment including nucleotide 590 to nucleotide 1078 of SEQ ID NO: 1 and oligonucleotides ZC16,421 (SEQ ID NO: 19) and ZC16,425 (SEQ ID NO: 20) as primers and zCTGF4 as template. The PCR reactions were run as follows: 1 cycle at 94°C for 1.5 minutes; 3 cycles of 94°C for 15 seconds; 50°C for 30; 72°C for 30 seconds; 12 cycles of 94°C for 15 seconds; 50°C for 20 seconds; 72°C for 30 seconds and 1 cycle at 72°C for 2 minutes. The PCR products were gel purified and mixed together as the template for the following PCR reaction: 1 cycle at 94°C for 1.5 minutes; 3 cycles of 94°C for 15 seconds; 54°C for 20; 68°C for 45 seconds; 20 cycles of

94°C for 15 seconds; 68°C for 45 seconds; and 1 cycle at 72°C for 2 minutes. This PCR product, which contained the entire polynucleotide sequence encoding for the mature zCTGF4 polypeptide, was gel purified and restriction 5 digested with BamH I and Xho I for use with the N- terminally tagged vector, pZP9NEE, described above. The zCTGF4 DNA sequence was ligated into the vector pZP9NEE and *E. coli* transformants selected, and designated pZP9NEE/zCTGF4.

10 Plasmid DNA was isolated and the region of the plasmid containing polynucleotides encoding for the tPA leader followed by the DNA encoding the GluGlu tag (SEQ ID NO: 16) and mature CTGF4 polypeptide were excised using a 5' EcoR I and a 3' XbaI site. The insert was sequence 15 analyzed for verification.

A similar cloning method was used to generate a C-terminally tagged expression construct. This construct, designated pZP9CEE/zCTGF4 comprises the pZP9CEE vector with DNAs encoding for the native zCTGF4 secretory signal 20 sequence, the mature zCTGF4 polypeptide (shown in SEQ ID NO: 1) and a C-terminal GluGlu tag (SEQ ID NO: 16).

The fragment from pZP9NEE/zCTGF4 containing DNA encoding for the the tPA secretory signal peptide, the GluGlu tag and the mature zCTGF4 were ligated into a 25 baculovirus vector designated pZBV4L. pZBV4L is a baculovirus expression vector derived from the FASTBAC vector of the Bac-to-Bac™ system (GIBCO-BRL, Gaithersburg, MD) and described in Luckow et al., J. Virol. 67:4566-4579, 1993. The pFASTBAC vector was 30 modified by removing the polyhedrin promoter and substituting the baculovirus basic protein promoter (Hill-Perkins et al., J. Gen. Virol. 71:971-976, 1990; Bonning et al., J. Gen. Virol. 75:1551-1556, 1994; and Chazenbalk et al., J. Biol. Chem. 270:1543-1549, 1995).

35 One microliter of zCTGF4NEE/pZBV4L expression construct is used to transform 20 µl DH10Bac (GIBCO-BRL,

Gaithersburg, MD) in 980 μ l SOC (2% Bacto Tryptone, 0.5% Bacto Yeast Extract, 10 ml 1M NaCl, 1.5 mM KCl, 10 mM MgCl₂, 10 mM MgSO₄ and 20 mM glucose), according to the manufacturer's specifications. The cells are incubated 48 hours at 37°C and 2 colonies in which virus had incorporated the plasmid (referred to as a "bacmid") were identified (white in color), and are isolated. Bacmid DNA is isolated and used to transfect *Spodoptera frugiperda* (Sf9) cells using transfection liposomes from Cellfectin (GIBCO-BRL). The cells are cultured at 27°C in shake flasks using 50-100 ml of SF900II serum free medium (GIBCO-BRL). After 3-4 days medium conditioned by the virus is harvested and used to infect Sf9 cells in mid-log growth at approximately 1E6 cells/ml.

The cells are scaled up by adding cell cultures to volumes of 15 liter when cells have achieved a density between 1-2E6 cell/ml and then infected with an MOI of 1-3. After 2 days at 27°C, the medium containing the protein and virus is harvested.

20

Example 5

A. Affinity Tagged zCTGF4 Protein
zCTGF4 expressed with an N-terminal or C-terminal GluGlu (EE) tag is purified as follows: A mixture of protease inhibitors is added to a 2000 ml sample of conditioned media from baculovirus-infected Sf9 cells to final concentrations of 2.5 mM ethylenediaminetetraacetic acid (EDTA, Sigma Chemical Co. St. Louis, MO), 0.001 mM leupeptin (Boehringer-Mannheim, Indianapolis, IN), and 0.001 mM pepstatin (Boehringer-Mannheim) and 0.4 mM Pefabloc (Boehringer-Mannheim). The sample is centrifuged at 10,000 rpm for 30 min at 4°C in a Beckman JLA-10.5 rotor (Beckman Instruments, Palo Alto, CA) in a Beckman Avanti J25I centrifuge (Beckman Instruments) to remove cell debris. To the supernatant fraction is added a 50.0 ml sample of anti-EE Sepharose,

prepared as described below, and the mixture is gently agitated on a Wheaton (Millville, NJ) roller culture apparatus for 18.0 h at 4°C.

The mixture is poured into a 5.0 x 20.0 cm
5 Econo-Column (Bio-Rad, Laboratories, Hercules, CA) and the gel is washed with 30 column volumes of phosphate buffered saline (PBS). The unretained flow-through fraction is discarded. When the absorbance of the effluent at 280 nM is less than 0.05, flow through the column is reduced to
10 zero and the anti-EE Sepharose gel is washed with 2.0 column volumes of PBS containing 0.2 mg/ml of EE peptide (AnaSpec, San Jose, CA). The peptide used has the sequence GluTyrMetGlu (SEQ ID NO: 16). After 1.0 h at 4°C, flow is resumed and the eluted protein is collected.
15 This fraction is the peptide elution. The anti-EE Sepharose gel is washed with 2.0 column volumes of 0.1M glycine, pH 2.5, and the glycine wash is collected separately. The pH of the glycine-eluted fraction is adjusted to 7.0 by the addition of a small volume of 10X
20 PBS and stored at 4°C.

The peptide elution is concentrated to 5.0 ml using a 15,000 molecular weight cutoff membrane concentrator (Millipore, Bedford, MA), according to the manufacturer's instructions. The concentrated peptide
25 elution is separated from free peptide by chromatography on a 1.5 x 50 cm Sephadex G-50 (Pharmacia, Piscataway, NJ) column equilibrated in PBS at a flow rate of 1.0 ml/min using a BioCad Sprint HPLC (PerSeptive BioSystems, Framingham, MA). Two-ml fractions are collected and the
30 absorbance at 280 nM is monitored. The first peak of material absorbing at 280 nM and eluting near the void volume of the column is collected. This fraction is pure N-terminally tagged zCTGF4 or C-terminally tagged zCTGF4. The pure material is concentrated as described above,
35 analyzed by SDS-PAGE and Western blotting with antiEE antibodies, aliquoted, and stored at -80°C.

Preparation of anti-EE Sepharose is done as follows: A 100 ml bed volume of protein G-Sepharose (Pharmacia, Piscataway, NJ) is washed 3 times with 100 ml of PBS containing 0.02% sodium azide using a 500 ml
5 Nalgene 0.45 micron filter unit. The gel is washed with 6.0 volumes of 200 mM triethanolamine, pH 8.2 (TEA, Sigma, St. Louis, MO), and an equal volume of EE antibody solution containing 900 mg of antibody is added. After an overnight incubation at 4°C, unbound antibody is removed
10 by washing the resin with 5 volumes of 200 mM TEA as described above. The resin is resuspended in 2 volumes of TEA, transferred to a suitable container, and dimethylpimelimidate-2HCl (Pierce, Rockford, IL), dissolved in TEA, is added to a final concentration of 36
15 mg/ml of gel. The gel is rocked at room temperature for 45 min and the liquid is removed using the filter unit as described above. Nonspecific sites on the gel are then blocked by incubating for 10 min. at room temperature with 5 volumes of 20 mM ethanolamine in 200 mM TEA. The gel is
20 then washed with 5 volumes of PBS containing 0.02% sodium azide and stored in this solution at 4°C.

B. Untagged zCTGF4 Protein

25 Protease inhibitors are added to the conditioned media of baculovirous-infected Sf9 cells and the media will be centrifuged as described above for the EE-tagged proteins. The supernatant fraction is applied to a 50.0 ml column of POROS HE1 (PerSeptive BioSystems, Framingham, MS) pre-equilibrated in 20 mM Tris-HCl, 50 mM NaCl, pH 7.4 at a flow rate of 2.0 ml/min with in-line dilution (three-fold final dilution) with water as diluent using a BioBad Sprint HPLC (PerSeptive BioSystems, Framingham, MS). Heparin-bound proteins are eluted with a 0.1-1.0 M
30 gradient of NaCl. Protein-containing fractions are identified by absorbance at 280 nM and by SDS-PAGE. Fractions containing zCTGF4 are identified by a band on SDS-PAGE gels of the appropriate molecular weight for
35

glycosylated zCTGF4 of approximately 40 kDa. The heparin-bound zCTGF4 pool is concentrated, applied to a Sephadex-G50 or a -G100 column, and eluted as described above. Purified zCTGF4 is characterized by SDS-PAGE, amino acid analysis, and N-terminal sequencing.

Example 6

A. Adenoviral Expression of zCTGF4

The protein coding region of zCTGF4 was amplified by PCR using primers that added FseI and AscI restriction sites at the 5' and 3' termini respectively. PCR primers ZC17948 (SEQ. ID. NO: 29) and ZC17949 (SEQ ID NO: 30) were used with a template containing the full-length zCTGF4 cDNA in a PCR reaction as follows: one cycle at 95°C for 5 minutes; followed by 15 cycles at 95°C for 1 min., 58°C for 1 min., and 72°C for 1.5 min.; followed by 72°C for 7 min.; followed by a 4°C soak. The PCR reaction product was loaded onto a 1.2 % (low melt) SeaPlaque GTG (FMC, Rockland, ME) gel in TAE buffer. The zCTGF4 PCR product was excised from the gel and purified using the QIAquick™ PCR Purification Kit gel cleanup kit as per kit instructions (Qiagen). The PCR product was then digested with FseI-AscI, phenol/chloroform extracted, EtOH precipitated, and rehydrated in 20ml TE (Tris/EDTA pH 8). The 1065 bp zCTGF4 fragment was then ligated into the FseI-AscI sites of the transgenic vector pTG12-8 (See, description herein) and transformed into DH10B competent cells by electroporation. Clones containing zCTGF4 were identified by plasmid DNA miniprep followed by digestion with FseI-AscI. A positive clone was confirmed by direct sequencing.

B. Preparation of DNA construct for generation of Adenovirus

The 1065 bp zCTGF4 cDNA was released from a TG12-8 vector using FseI and AscI enzymes. The cDNA was

isolated on a 1% low melt SeaPlaque GTG™ (FMC, Rockland, ME) gel, and was then excised from the gel. The gel slice was melted at 70°C, extracted twice with an equal volume of Tris buffered phenol, and EtOH precipitated. The DNA 5 was resuspended in 10µl H₂O.

The zCTGF4 cDNA was cloned into the FseI-AscI sites of a modified pAdTrack CMV (He et al., PNAS 95:2509-2514, 1998). This construct contains the GFP marker gene. The CMV promoter driving GFP expression was replaced with 10 the SV40 promoter and the SV40 polyadenylation signal was replaced with the human growth hormone polyadenylation signal. In addition, the native polylinker was replaced with FseI, EcoRV, and AscI sites. This modified form of pAdTrach CMV was named pZyTrack. Ligation was performed 15 using the Fast-Link™ DNA ligation and screening kit (Epicentre Technologies, Madison, WI). In order to linearize the plasmid, approximately 5 µg of the pZyTrack zctgf4 plasmid was digested with PmeI. Approximately 1 µg of the linearized plasmid was cotransformed with 200ng of 20 supercoiled pAdEasy (He et al., supra.) into BJ5183 cells. The co-transformation was done using a Bio-Rad Gene Pulser at 2.5kV, 200 ohms and 25mFa. The entire co-transformation was plated on 4 LB plates containing 25 µg/ml kanamycin. The smallest colonies were picked and 25 expanded in LB/kanamycin and recombinant adenovirus DNA identified by standard DNA miniprep procedures. Digestion of the recombinant adenovirus DNA with FseI-AscI confirmed the presence of zCTGF4. The recombinant adenovirus miniprep DNA was transformed into DH10B competent cells 30 and DNA prepared using a Qiagen maxi prep kit as per kit instructions.

C. Transfection of 293A Cells with Recombinant DNA

Approximately 5 µg of recombinant adenoviral DNA 35 was digested with PacI enzyme (New England Biolabs) for 3 hours at 37°C in a reaction volume of 100 µl containing

20-30U of PacI. The digested DNA was extracted twice with an equal volume of phenol/chloroform and precipitated with ethanol. The DNA pellet was resuspended in 10 μ l distilled water. A T25 flask of QBI-293A cells (Quantum 5 Biotechnologies, Inc. Montreal, QC. Canada), inoculated the day before and grown to 60-70% confluence, were transfected with the PacI digested DNA. The PacI-digested DNA was diluted up to a total volume of 50 μ l with sterile HBS (150mM NaCl, 20mM HEPES). In a separate tube, 20 μ l 10 DOTAP (Boehringer Mannheim, 1mg/ml) was diluted to a total volume of 100 μ l with HBS. The DNA was added to the DOTAP, mixed gently by pipeting up and down, and left at room temperature for 15 minutes. The media was removed from the 293A cells and washed with 5 ml serum-free MEM-alpha 15 (Gibco BRL) containing 1 mM Sodium Pyruvate (GibcoBRL), 0.1 mM MEM non-essential amino acids (GibcoBRL) and 25 mM HEPES buffer (GibcoBRL). 5 ml of serum-free MEM was added to the 293A cells and held at 37°C. The DNA/lipid mixture was added drop-wise to the T25 flask of 293A cells, mixed 20 gently and incubated at 37°C for 4 hours. After 4 h the media containing the DNA/lipid mixture was aspirated off and replaced with 5 ml complete MEM containing 5% fetal bovine serum. The transfected cells were monitored for Green Fluorescent Protein (GFP) expression and formation 25 of foci, i.e., viral plaques.

Seven days after transfection of 293A cells with the recombinant adenoviral DNA, the cells expressed the GFP protein and started to form foci. These foci are viral "plaques" and the crude viral lysate was collected 30 by using a cell scraper to collect all of the 293A cells. The lysate was transferred to a 50ml conical tube. To release most of the virus particles from the cells, three freeze/thaw cycles were done in a dry ice/ethanol bath and a 37° waterbath.

D. Amplification of Recombinant Adenovirus (rAdV)

The crude lysate was amplified (Primary (1°) amplification) to obtain a working "stock" of zCTGF4 rAdV lysate. Ten 10cm plates of nearly confluent (80-90%) 293A cells were set up 20 hours previously, 200ml of crude rAdV lysate added to each 10cm plate and monitored for 48 to 72 hours looking for CPE under the white light microscope and expression of GFP under the fluorescent microscope. When all of the 293A cells showed CPE (Cytopathic Effect) this 1° stock lysate was collected and freeze/thaw cycles performed as described under Crude rAdV Lysate.

Secondary (2°) Amplification of zCTGF4 rAdV was obtained as follows: Twenty 15 cm tissue culture dishes of 293A cells were prepared so that the cells were 80-90% confluent. All but 20 mls of 5% MEM media was removed and each dish was inoculated with 300-500 ml 1° amplified rAdV lysate. After 48 hours the 293A cells were lysed from virus production and this lysate was collected into 250 ml polypropylene centrifuge bottles and the rAdV purified.

20

E. AdV/cDNA Purification

NP-40 detergent was added to a final concentration of 0.5% to the bottles of crude lysate in order to lyse all cells. Bottles were placed on a rotating platform for 10 min. agitating as fast as possible without the bottles falling over. The debris was pelleted by centrifugation at 20,000 X G for 15 minutes. The supernatant was transferred to 250 ml polycarbonate centrifuge bottles and 0.5 volumes of 20% PEG8000/2.5M NaCl solution added. The bottles were shaken overnight on ice. The bottles were centrifuged at 20,000 X G for 15 minutes and supernatant discarded into a bleach solution. The white precipitate in two vertical lines along the wall of the bottle on either side of the spin mark is the precipitated virus/PEG. Using a sterile cell scraper, the precipitate from 2 bottles was resuspended in 2.5 ml PBS.

The virus solution was placed in 2 ml microcentrifuge tubes and centrifuged at 14,000 X G in the microfuge for 10 minutes to remove any additional cell debris. The supernatant from the 2ml microcentrifuge tubes was 5 transferred into a 15ml polypropylene snapcap tube and adjusted to a density of 1.34 g/ml with cesium chloride (CsCl). The volume of the virus solution was estimated and 0.55 g/ml of CsCl added. The CsCl was dissolved and 1 ml of this solution weighed 1.34 g. The solution was 10 transferred polycarbonate thick-walled centrifuge tubes 3.2ml (Beckman) and spin at 80,000rpm (348,000 X G) for 3-4 hours at 25°C in a Beckman Optima TLX microultracentrifuge with the TLA-100.4 rotor. The virus formed a white band. Using wide-bore pipette tips, the 15 virus band was collected.

The virus from the gradient has a large amount of CsCl which must be removed before it can be used on cells. Pharmacia PD-10 columns prepacked with Sephadex G-25M (Pharmacia) were used to desalt the virus preparation. 20 The column was equilibrated with 20 ml of PBS. The virus was loaded and allowed to run into the column. 5 ml of PBS was added to the column and fractions of 8-10 drops collected. The optical densities of 1:50 dilutions of each fraction was determined at 260 nm on a 25 spectrophotometer. A clear absorbance peak was present between fractions 7-12. These fractions were pooled and the optical density (OD) of a 1:25 dilution determined. A formula is used to convert OD into virus concentration: (OD at 260nm) (25) (1.1×10^{12}) = virions/ml. The OD of a 30 1:25 dilution of the zctgf4 rAdV was 0.221, giving a virus concentration of 6×10^{12} virions/ml.

To store the virus, glycerol was added to the purified virus to a final concentration of 15%, mixed gently but effectively, and stored in aliquots at -80°C.

F. Tissue Culture Infectious Dose at 50% CPE (TCID 50)
Viral Titration Assay

A protocol developed by Quantum Biotechnologies, Inc. (Montreal, Qc. Canada) was followed to measure recombinant virus infectivity. Briefly, two 96-well tissue culture plates were seeded with 1×10^4 293A cells per well in MEM containing 2% fetal bovine serum for each recombinant virus to be assayed. After 24 hours 10-fold dilutions of each virus from 1×10^{-2} to 1×10^{-14} were made in MEM containing 2% fetal bovine serum. $100\mu\text{l}$ of each dilution was placed in each of 20 wells. After 5 days at 37°C , wells were read either positive or negative for Cytopathic Effect (CPE) and a value for "Plaque Forming Units/ml" (PFU) is calculated.

TCID₅₀ formulation used was as per Quantum Biotechnologies, Inc., above. The titer (T) is determined from a plate where virus used is diluted from 10^{-2} to 10^{-14} , and read 5 days after the infection. At each dilution a ratio (R) of positive wells for CPE per the total number of wells is determined.

To Calculate titer of the undiluted virus sample: the factor, "F" = $1 + d(S - 0.5)$; where "S" is the sum of the ratios (R); and "d" is Log₁₀ of the dilution series, for example, "d" is equal to 1 for a ten-fold dilution series. The titer of the undiluted sample is $T = 10^{(1+F)} = \text{TCID}_{50}/\text{ml}$. To convert TCID₅₀/ml to pfu/ml, 0.7 is subtracted from the exponent in the calculation for titer (T). The zCTGF4 adenovirus had a titer of 7.1×10^{10} pfu/ml.

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Example 7

Transgenic Expression

Transgenic animals expressing zCTGF4 genes were made using adult, (B6D2f1, 2-8 months, (Taconic Farms)), prepubescent fertile females (donors) (B6C3f1, 4-5 weeks,

(Taconic Farms)) and adult fertile females (B6D2f1, 2-4 months, (Taconic Farms) as parents.

The donors were injected with approximately 8 IU/mouse of Pregnant Mare's Serum gonadotrophin (Sigma, 5 St. Louis, MO) I.P., and 46-47 hours later, 8 IU/mouse of human Chorionic Gonadotropin (hCG (Sigma)) were administered I.P. to induce superovulation.

Fertilized eggs were collected and stored in a 37°C/5% CO₂ incubator until microinjection.

10 10-20 micrograms of plasmid DNA containing a cDNA of the zCTGF4 gene was linearized, gel-purified, and resuspended in 10 mM Tris pH 7.4, 0.25 mM EDTA pH 8.0, at a final concentration of 5-10 nanograms per microliter for microinjection.

15 Plasmid DNA was microinjected into harvested eggs and were penetrated with an injection needle, into one or both of the haploid pronuclei.

20 The following day 2-cell embryos were transferred into pseudopregnant recipients. The recipients were returned to cages in pairs, and allowed 19-21 days gestation. After birth, 19-21 days postpartum was allowed before weaning. The 25 weanlings were sexed and placed into separate sex cages, and a 0.5 cm biopsy (used for genotyping) was snipped off the tail with clean scissors.

25 Genomic DNA was prepared from the tail snips using a Qiagen Dneasy kit following the manufacturer's instructions. Genomic DNA was analyzed by PCR using primers designed to the human growth hormone (hGH) 3' UTR portion of the transgenic vector. A region unique to the 30 human sequence was identified from an alignment of the human and mouse growth hormone 3' UTR DNA sequences, ensuring that the PCR reaction does not amplify the mouse sequence. Primers zc17251 (SEQ ID NO: 31) and zc17252 (SEQ ID NO: 32) amplify a 368 base pair fragment of hGH. 35 In addition, primers zc17156 (SEQ ID NO: 33) and zc17157 (SEQ ID NO: 34), which hybridize to vector sequences and

amplify the cDNA insert, were often used along with the hGH primers. In these experiments, DNA from animals positive for the transgene generated two bands, a 368 base pair band corresponding to the hGH 3' UTR fragment and a 5 band of variable size corresponding to the cDNA insert.

Once 9 animals were confirmed to be transgenic (TG), they were back with C57B1/6 wild-type mates. As pups were born and weaned, the sexes were separated, and their tails snipped for genotyping.

10 To check for expression of a transgene in a live animal, a small partial hepatic biopsy was collected. The collected liver biopsy was transferred to a 14 ml polypropylene round bottom tube and snap frozen in liquid nitrogen and then stored on dry ice.

15 Analysis of the mRNA expression level of each transgene was done using an RNA solution hybridization assay. In addition, transgenic mice were observed to be smaller than wild-type mice from the same litter. Thymus and spleen weights were observed to be lower when 20 normalized by brain-to-organ weights. Histological examination revealed that most of the transgenic mice had some pancreatic atrophy and some cardiomyopathy.

From the foregoing, it will be appreciated that, 25 although specific embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the invention is not limited except as by the appended claims.